1	Human health risks for <i>Legionella</i> and <i>Mycobacterium</i>
2	avium complex (MAC) from potable and non-potable uses
3	of roof-harvested rainwater
4	
5	
6	Kerry A. Hamilton ^{a,b,*} , Warish Ahmed ^a , Simon Toze ^a , Charles N. Haas ^b
7	
8	
9	
10	
11	^a CSIRO Land and Water, Ecosciences Precinct, 41 Boggo Road, Qld 4102, Australia
12	^b Drexel University, 3141 Chestnut Street, Philadelphia, PA 19104, USA
13	
14	
15	
16	
17	Running title: QMRA of opportunistic pathogens in roof-harvested rainwater
18	
19	XC
20	
21	
22	
23	
24	
25	
26	
27	
28 29	* Corresponding author. Kerry Hamilton. Mailing address: Drexel University Department of Civil, Architectural, and Environmental Engineering, 3141 Chestnut Street, Philadelphia, Pennsylvania,

30 19104, USA. Tel.: +1 215 895 2000; Fax: +1 215 895 1363. E-mail address: <u>kh495@drexel.edu</u>.

31 ABSTRACT

A quantitative microbial risk assessment (QMRA) of opportunistic pathogens Legionella pneumophila (LP) and Mycobacterium avium complex (MAC) was undertaken for various uses of roof-harvested rainwater (RHRW) reported in Brisbane, Australia to identify appropriate usages and guide risk management practices. Risks from inhalation of aerosols due to showering, swimming in pools topped up with RHRW, use of a garden hose, car washing, and toilet flushing with RHRW were considered for LP while both ingestion (drinking, produce consumption, and accidental ingestion from various activities) and inhalation risks were considered for MAC. The drinking water route of exposure presented the greatest risks due to cervical lymphadenitis and disseminated infection health endpoints for children and immune-compromised populations, respectively. It is therefore not recommended that these populations consume untreated rainwater. LP risks were up to 5 orders of magnitude higher than MAC risks for the inhalation route of exposure for all scenarios. Both inhalation and ingestion QMRA simulations support that while drinking, showering, and garden hosing with RHRW may present the highest risks, car washing and clothes washing could constitute appropriate uses of RHRW for all populations, and toilet flushing and consumption of lettuce irrigation with RHRW would be appropriate for non- immune-compromised populations. Keywords: Roof-harvested rainwater; opportunistic pathogens; guantitative microbial risk assessment (QMRA); Legionella pneumophila; Mycobacterium avium complex.

59 **1. Introduction**

60 Globally, harvested rainwater is used to supplement both potable and non-potable water supplies. 61 In Australia, roof-harvested rainwater (RHRW) constitutes an important source of water for many 62 households; in 2010, 32% of Australian households had a rainwater tank and rainwater tanks were 63 the main source of drinking water for 13.6% of Queensland households (ABS, 2010). Queensland 64 rainwater tank owners have reported numerous potable and non-potable uses for their RHRW, 65 including drinking, cooking, clothes washing, showering, pool top-up, gardening, car washing, 66 ornamental water features, toilet flushing, filling fish tanks, and pet washing (Hamilton et al., 2016). 67 This indicates the potential for exposure to rainwater through numerous scenarios. 68 Previous studies have identified high concentrations of both enteric pathogens such as 69 Salmonella spp., E. coli, Campylobacter spp., Cryptosporidium spp., and Giardia spp. (Ahmed et al., 70 2011; Ahmed et al., 2010; Crabtree 1996) and opportunistic pathogens such as Legionella 71 pneumophila (LP), Mycobacterium avium complex (MAC), Aeromonas hydrophila, Staphylococcus 72 aureus, Pseudomonas aeruginosa, and Acanthamoeba spp. (Ahmed et al., 2014; Hamilton et al., 73 2016) in subtropical rainwater tanks. Opportunistic pathogens cause illness primarily in individuals 74 with underlying health conditions, children, and/or the elderly. However, they are a growing cause of 75 drinking water-associated disease outbreaks worldwide (Falkinham 3rd et al., 2015). 76 There is epidemiologic evidence of disease cases associated with enteric pathogens (Brodribb et 77 al., 1995; Franklin et al., 2009; Koplan, 1978; Merritt et al., 1999; Murrell and Stewart, 1983; Simmons 78 and Smith 1997) as well as opportunistic pathogens LP (Schlech III et al., 1985; Simmons et al., 79 2008) and MAC (Lumb et al., 2004) in RHRW. While drinking water guidelines used to determine the 80 safety of Australian rainwater specify the non-detection of E. coli in 100 mL of water (NHMRC-81 NRMMC, 2011; WHO, 2004), there is no consensus about which end-uses of rainwater are 82 appropriate with regards to opportunistic pathogen associated health risks. No study has assessed 83 the full suite of potential rainwater uses for an opportunistic pathogen to make such a determination. 84 This is especially important as treatment options are limited for rainwater tank owners and are typically limited to gutter protection, first-flush devices, or point-of-use filters. These options have 85 86 limited efficacy for removing pathogens (Dobrowsky et al., 2015a; Dobrowsky et al., 2015b; 87 Egodawatta et al., 2009; Jordan, 2008; Kus et al., 2010; Mendez et al., 2011; Reyneke et al., 2016).

88 Quantitative microbial risk assessment (QMRA) can be used for the purposes of estimating the 89 human health risk associated with exposure to pathogens in environmental matrices using a process 90 of hazard identification, exposure assessment, dose response, and risk characterization (Haas et al., 91 1999). Due to their direct linkage to RHRW-associated disease cases (Lumb et al., 2004; Simmons et 92 al., 2008), global epidemiologic importance (Falkinham 3rd et al., 2015), and known occurrence in 93 RHRW (Hamilton et al., 2016), LP and MAC were chosen as index opportunistic pathogens for a 94 QMRA estimate. While there are over 50 species of Legionella and several are human pathogens, LP 95 is the most common species that causes the severe pneumonia-like illness Legionnaires' Disease, as 96 well as the less severe form of illness, Pontiac fever (Diederen, 2008; Muder and Victor, 2002). The 97 rate of Legionellosis in Australia was 13 per million people in 2012, and eighty cases were reported in 98 Queensland in 2015 (Australian Government Department of Health, 2016; Phin et al., 2014). MAC is a 99 subset of non-tuberculous mycobacteria (NTM) that can cause skin and soft tissue infections or 100 cervical lymphadenitis in immune-competent patients, disseminated infections in immune-101 compromised patients, and pulmonary disease in both healthy and immune-compromised groups (Falkinham 3rd, 1996). MAC were the most common pathogen in NTM isolates in Queensland in 2005, 102 103 and most frequently identified isolate in NTM cases in the Northern Territory, Australia from 1989-104 1997 (O'Brien et al., 2000; Thomson et al., 2013), however NTM-associated illnesses are not 105 reportable in Australia. MAC is comprised of 9 species (M. avium, M. intracellulare, M. arosiense, M. 106 chimaera, M. colombiense, M. marseillense, M. timonense, M. bouchedurhonense, and M. ituriense) 107 (Falkinham 3rd, 2013). The most human-relevant species and therefore the focus of this QMRA are *M*. 108 avium (comprised of four subspecies: paratuberculosis, avium, hominissuis, and silvaticum), M. 109 intracellulare, and M. chimaera (Hamilton et al., 2017).

110 Previous QMRA studies of RHRW have focused on enteric pathogens or LP, typically focusing on 111 one or two exposure scenarios (Ahmed et al., 2010; de Man et al., 2014a; Fewtrell and Kay, 2007; 112 Lim et al., 2015; Lim and Jiang, 2013; Schoen and Garland, 2015; Schoen et al., 2014). This has 113 been partially due to the lack of dose response models for opportunistic pathogens such as MAC. A single previous assessment of exposure to MAC is available for treated tap water from a centralized 114 115 distribution system (Rice et al., 2005). However, this study did not quantify health risks. A dose 116 response model has since been developed for one MAC subspecies, *M. avium* subsp. 117 paratuberculosis (MAP) (Breuninger and Weir, 2015), however the relationship between human

exposure to this subspecies and the development of health effects [it is postulated that Crohn's
disease may be the health outcome for this pathogen (Pierce, 2009; Waddell et al., 2015)] is
contentious and is therefore excluded from the current analysis. A family of MAC dose response
models was recently developed for human-relevant species of MAC in environmental matrices,
allowing for development of a population- and exposure-route specific QMRA for MAC risks (Hamilton

123 et al., 2017). For LP, generally only inhalation or aspiration routes are considered relevant, with

inhalation being the most common exposure route (Ellis, 1993).

125 The goals of the current study are therefore to assess the health risks from index pathogens LP

and MAC by conducting a QMRA of multiple potential exposure scenarios with (1) inhalation exposure

to LP and (2) inhalation and ingestion exposures to MAC.

128

129 2. Materials and Methods

130 2.1 Exposure models

131 In our previous study, Queensland rainwater tank owners reported using RHRW for drinking, cooking, 132 clothes washing, showering, pool top-up, gardening, car washing, ornamental water features, toilet 133 flushing, filling fish tanks, and pet washing (Hamilton et al., 2016) (Supplementary Table S1). In that 134 study, LP and MAC were measured in 134 rainwater tanks from Brisbane, Australia. The use of RHRW for ornamental water features, filling fish tanks, and pet washing were reported by less than 135 136 2% of surveyed residents, and were therefore excluded from the current QMRA analysis. The 137 remaining potential exposure pathways for LP and MAC are shown in Fig. 1. As reported in a 138 previous study (Hamilton et al., 2017), exposure to MAC species can occur through: (1) the ingestion 139 of food, water, or soil; (2) inhalation of water aerosols or soil dusts; (3) aspiration; or (4) iatrogenic 140 exposure. Scenarios (1) and (2) were considered here as it is unlikely to use harvested rainwater in a 141 medical setting, and limited information is available regarding aspiration rates and the amount of 142 pathogenic material transferred to the lungs during each aspiration event. Exposure models for each 143 scenario are summarized below and in Fig. 1.

144 It is noted that in addition to the importance of the exposure route, MAC is associated with 145 different health endpoints depending on the population exposed (Hamilton et al. 2017). Ingestion 146 routes of exposure are associated with cervical lymphadenitis in children or disseminated infection in 147 immune-deficient individuals (Falkinham 2013). Immune-deficient individuals can be either adults or 148 children. Children over the age of 12 are rarely affected by MAC lymphadenitis, except when 149 immunodeficiencies or disseminated disease are present (Lai et al. 1984), and cervical lymphadenitis 150 is most common in children from ages 1 to 5 (Inderlied et al. 1993, Haverkamp et al. 2004, Wolinsky 151 1995, Lincoln et al, 1972, Hazra et al. 1999, Tebruegge et al. 2016). For this reason, MAC risks for 152 the ingestion pathway were calculated separately for a general immune-deficient population and 153 children ages 1-2, 3-5, and 6-12 based on differences in disease presentation, susceptibility to MAC, 154 and exposure patterns. Despite similar child susceptibility to MAC from age 1 to 5, activity patterns 155 were different depending on ages and thus two categories (1-2 and 3-5) were used. Inhalation 156 exposures to MAC have been shown to be epidemiologically important for healthy, post-menopausal 157 females or older males with pre-existing lung conditions and/or risk factors, however healthy 158 individuals may also become ill (Field et al. 2004). Children may also be affected via the inhalation 159 route, although this is less common (Freeman et al. 2009, Nolt et al. 2003). Additionally, skin and 160 antibody test reactivity studies have indicated increasing levels of exposure with age and evidence of asymptomatic infection with MAC (Griffith et al. 2007). However, sufficient information is not available 161 at this time to develop separate risk expressions for MAC respiratory infection on the basis of specific 162 163 age range or susceptibility factors. Therefore, for the inhalation models, MAC infection risks were 164 calculated for a general population rather than age- or immune status- separated populations. 165 Legionnaires' disease has been reported in children (Luttichau et al. 1998; Aebischer et al. 1999; Campins et al. 2000), and a general population at risk was also considered. 166 2.2 Ingestion exposure models 167

As Legionellosis is contracted through inhalation or aspiration, ingestion exposure models apply only to MAC. The general process for ingestion of RHRW for a variety of applications for MAC can be described by equation (1). Additional factors and modifications for each scenario are described below and summarized in Table 2.

$$D_{i,j} = \frac{1}{R} C_{RHRW,MAC} V_{ing,j} \tag{1}$$

173

172

Where $D_{i,j}$ = The daily dose of pathogen *i* (where *i* = LP or MAC) for exposure scenario *j* (where *j* = drinking, eating produce, etc.), C_{RHRW} = Concentration of MAC in RHRW, *R* = recovery efficiency (%), and V_{ing} = the volume of RHRW ingested per exposure event.

177 2.2.1 Drinking

A mean of 0.869 L per day and 95th percentile 2.717 L per day (US EPA, 2011a) was used to 178 179 construct a lognormal distribution for daily drinking water intake for a general population. For children, 180 drinking water exposure parameters reported for ages 1 to <2 years, 3 to <6 years, and 6 to <11 181 years were used for 1-2, 3-5, and 6-11 year-old child age groups, respectively (US EPA, 2011a). 182 Parameters reported for ages 1 to <2 years and 2 to <3 years were used to fit overall 1 to 2 year 183 parameters by generating 10,000 observations from each set of parameters (1 to <2 year and 2 to <3 184 year) which were then concatenated. The age brackets were weighted equally as the proportion of 1 185 and 2 year olds in the Queensland population was equivalent in 2015 (ABS, 2016). A new distribution 186 was fit to the combined data.

187 Two scenarios, without filtration and with filtration (equation 2) were considered as some 188 participants in our previous rainwater study (Hamilton et al., 2016) had under-sink point-of-use (POU) 189 filters. Chlorine disinfection was not considered as only one participant of 134 households reported 190 having ever used chlorine to disinfect their rainwater (Hamilton et al., 2016). The most common treatment method for those who reported treating their rainwater was filtration (29/134 participants) 191 192 (Hamilton et al., 2016). For a POU filter used with rainwater systems, a previous study reported a 193 39% (0.4 log) reduction in *E. coli* bacteria (Jordan, 2008). A sand filter used with rainwater tanks was 194 associated with a 99% (2 log) removal of bacteria (Ahammed and Meera, 2010). A uniform 195 distribution for filtration of 0.4-2 log removals was used.

$$D_{i,j} = \frac{1}{R} C_{RHRW,MAC} V_{ing,j} 10^{-L}$$
(2)

197 Where, L = the number of log removals for filtration.

198 2.2.2 Consumption of raw produce

199 The consumption of uncooked lettuce was chosen as the index scenario for eating produce irrigated 200 with RHRW. The selection of lettuce was due to its high water retention compared to other crops 201 because of its large and uneven surface area, its high probability of being eaten raw, high lettuce 202 consumption compared to other produce crops by Australian populations, and short shelf life that 203 bounds the possible time between harvest and consumption, especially in a subtropical climate 204 (Ahmed et al., 2016). Contamination with MAC in lettuce can occur through processes of: (1) the 205 irrigation water adheres to the outside of the plant and (2) MAC is internalized into plants (Kaevska et 206 al., 2014). Internalization can occur through uptake through the roots, or through stomata or wounds 207 present on the leaf surface (Hirneisen et al., 2012). These microorganisms would not be washed off

by the consumer as they are located inside the plant. To account for both of these processes
(surface-attached and internalized *M. avium*), the total dose of MAC on the surface of lettuce and the
MAC internalized in the plant are summed to arrive at the total dose (equation 3 - equation 5) (SalesOrtells et al., 2014).

212

213
$$D_{MAC,surface} = \frac{1}{R} C_{RHRW,MAC} V_R 10^{-k_{f,s}t_f} 10^{-W} I B_a F_{CL} \dots \dots \dots \dots \dots \dots \dots \dots (Eqn. 3)$$

214

215

$$D_{MAC,internalized} = \frac{1}{R} C_{RHRW,MAC} F_{int} V_R I B_a F_{CL} \dots \dots \dots \dots \dots \dots (Eqn. 4)$$
$$D_{MAC,total} = D_{MAC,surface} + D_{MAC,internalized} \dots \dots \dots \dots \dots \dots \dots \dots \dots (Eqn. 5)$$

Where V_R = volume retained per gram of lettuce during irrigation (L per gram), I_a = gram. 216 consumed per kg-person per day for age range a where a= ages 1-2, 3-5, 6-12, or a general 217 population (g/kg/day), B_a = body weight for each age range a of interest (kg), F_{CL} = fraction of 218 219 population of interest that consumes lettuce, $K_{f,s}$ = in-field decay constant for MAC on the surface of lettuce (per day), $W = \log$ reductions due to washing of lettuce prior to consumption, $K_{t,int} =$ in-field 220 decay constant for internalized MAC (per day), F_{int} = the internalized fraction of MAC in the irrigation 221 222 water that is found in the leaves (in CFU per gram of lettuce), t_f = withholding time between harvest 223 and consumption (days). Lettuce is assumed to be immediately consumed by the homeowner after 224 harvest, and, therefore decay of MAC due to transport and/or storage of the lettuce is not considered. 225 Internalization has typically been reported for enteric bacteria or viruses in a variety of produce (Dicaprio et al., 2012; Solomon et al., 2002; Wei et al., 2011), however, only one report of 226 227 internalization of *M. avium* in plants is available. Internalization rates were calculated from Kaevska et 228 al., (2014) (Fig. 2 and 3) data from field studies with tomato plant leaves by computing (concentration 229 recovered from leaves after surface washing) / (concentration of seeding inoculum) for lettuce. Internalized fractions ranged from 1.13×10^{-5} to 9.49×10^{-4} for *M. avium* subsp. *paratuberculosis* and 230 1.00×10^{-8} to 1.37×10^{-5} for *M. avium* subsp. *avium*. A uniform distribution using the more 231 conservative M. avium subsp. paratuberculosis values was used. During field experiments conducted 232 233 by Kaevska et al., (2014) over 8 weeks, concentrations of *M. avium* in leaves did not consistently increase or decrease between 3 (M. avium subsp. avium) or 4 (M. avium subsp. paratuberculosis) 234 235 sets of experiments. For this reason, decay was not included in the model for internalized *M. avium*. 236 For the volume of water retained by lettuce during irrigation, Shuval et al., (1997) estimated a 237 mean and standard deviation of 0.108 ± 0.019 mL per g for green oak lettuce planted in beds with

238 three staggered rows spaced 30 cm apart with fixed-set overhead sprinklers at 10 min intervals. It 239 was assumed that microorganisms are homogeneously distributed in RHRW stored in tanks and 240 100% of the organisms in the retained irrigation water become initially attached to the lettuce surface. 241 Decay experiments for *M. avium* on the surface of plants were not available, however Cook et al., 242 (2013) performed decay experiments for M. avium subsp. paratuberculosis exposed to silage 243 exudates derived from grass and alfalfa. A decay k value of -0.0484 per day was reported. Removal 244 of *E. coli* during washing with water of $0.3 \pm 0.1 \log$ (Holvoet et al., 2014) was assumed to be 245 representative of removal of *M. avium* and *M. intracellulare*. 246 A previously reported consumption distribution for the general population was used for lettuce

(Barker et al. 2013). Body weights for the general Australian population were computed by conducted
weighted sampling of distributions simulated from each age range reported in the literature based on
the age profile of the greater Brisbane area (ABS 1998, ABS 2016, CSIRO 2008). Australian data was
not available in a format that could be separated for ages 1-2, 3-5, and 6-12 years. For this reason,

lettuce consumption and body weight data from a US population was used for children 1-12 years old(USEPA 2011a).

253 2.2.3 Showering

A uniform distribution of water accidentally ingested per daily shower event was used of 58 μL - 1.9
 mL (Ahmed et al., 2010). Information was not available for child-specific exposure, therefore these
 estimates were used for both child (ages 6- 12) and general population exposures.

257 2.2.4 Garden hose use

258 Estimates for accidental ingestion during use of a garden hose has been reported to range from 0.002

259 -1.9 μL (Ahmed et al., 2010), up to 1 mL (NRMMC-EPHC-AHMC, 2006), or 1.1 mL (Schoen et al.,

260 2014). Irrigation of lettuce with recycled water was estimated to occur 90 times per year (NRMMC-

261 EPHC-AHMC, 2006). Gardening was not considered for child-specific exposure scenarios.

262 2.2.5 Car washing

263 The total volume of water consumed during 10 min of car washing using a high-pressure spray device

- was recently estimated using cyanuric acid as a tracer of water ingestion. Among 26 participants, the
- accidental ingestion volume per car washing event was estimated to range from 0.06 to 3.79 mL

266 (Sinclair et al., 2016). A monthly car washing frequency was assumed (Villarreal and Dixon, 2005).

267 Car washing was not considered for child-specific exposure scenarios.

268 2.2.6. Pool top-up and bathing

Australian pool-owners reported using RHRW as well as drinking water to fill their swimming pools and all pools observed in the previous study were outdoor family pools (Hamilton et al., 2016). The distribution of the proportion of drinking water: RHRW is not known as pool sizes, tank sizes, and pool maintenance practices vary considerably. As a result, scenarios of 10, 50, and 90% dilution of RHRW with opportunistic pathogen-free water in pools were modelled (equation 6). Additional pathogen decay through mixing of RHRW with tap water with a chlorine residual was considered to be minimal and potential opportunistic pathogen removal due to pool filters was not considered.

276

$$D_{i,j} = \frac{1}{R} C_{RHRW} V_{ing} D \tag{6}$$

277 Where D = dilution factor and D = 100% for the bathing water scenario (100% rainwater).

An ingestion volume of 51.5 mL and standard deviation of 103.5 mL per swim (lognormal parameters μ = 2.92 mL, σ = 1.43 mL) for a combined population of children and adults was used (Dufour et al., 2006). For child ingestion during swimming in pools or young child exposure during daily baths (ages 1 to 5; in place of showers), the distribution for child swimming pool ingestion from Schets et al. (2011) for Dutch children <15 years was used. Information was not available to fit separate distributions to the separate child age groups of interest.

284 Limited information is available regarding the frequency of swimming pool use and no studies 285 of swimming frequency from Australian freshwater pools could be found. Only the study from the Netherlands (Schets et al. 2011) reported swimming pool use frequency as 13-24 times per year on 286 287 average (95% CI up to 65 days per year for adults and 91 days per year for children) for a five-month 288 swimming season lasting from May 1 through October 1. Brisbane is coolest from June through 289 August (Australian Government Bureau of Meteorology, 2016). Assuming that the Queensland 290 swimming season takes place during a 9 month swimming season from September through May and 291 that the rate of Netherlands pool swimming frequency is similar, the distributions for swimming 292 frequency in swimming pools from Schets et al. were scaled to account for a 33% longer swimming 293 season in Australia. The child distribution (<15 years of age) was used for all child age brackets due 294 to lack of information on specific age ranges. To obtain parameters for a general swimming pool 295 frequency distribution, distributions were simulated from the male, female, and child swimming 296 frequency parameters (Schets et al. 2011) and weighted according to the portion of adults and 297 children < 15 years of age in the greater Brisbane population (ABS, 2016). The distributions were

298 concatenated and a negative binomial distribution was fit to the resulting dataset. Using the modified distributions, average and upper bound (95th percentile) point estimates for swimming frequency were 299 300 calculated.

301 2.2.7. Toilet flushing

302 For toilet flushing, estimates of the volume consumed per flush range from 0.01 mL (NRMMC-EPHC-

303 AHMC, 2006) to 0.3 mL (Schoen et al., 2014). A mean of 5 flushes per day was reported (Mayer and

- 304 DeOreo, 1999). Toilet flushing exposures were considered for child exposure models for ages > 3
- 305 years only as the average age of toilet training is approximately 3 years (Bloom et al. 1993). Child-
- 306 specific exposure information was not available, therefore the same distributions for number of
- 307 flushes were used for all age groups.

308 2.2.8. Clothes washing

An exposure volume of 0.01 mL 100 times per year is estimated for exposure to water used for 309

310 clothes washing (NRMMC-EPHC-AHMC, 2006). No aerosols were observed during clothes washing

311 experiments, therefore, only ingestion is considered for clothes washing (O'Toole et al., 2008a).

312 Clothes washing exposures were not considered for child-specific exposure models.

313 2.3. Inhalation exposure models

314 2.3.1 Showering, garden hosing, car washing, toilet flushing

Inhalation exposures were considered for both LP and MAC. However, the MAC health endpoints and 315 316 population at risk differs from ingestion exposures and as a result all inhalation models accounted for 317 pulmonary infection risks in a general population rather than adult- or child- specific pathways. LP 318 concentrations are parameterized slightly differently than MAC due to the small number of detects (n 319 = 4 positive out of 134 samples) in rainwater tanks in the study by Hamilton et al. (2016). Maximum 320 likelihood used for MAC is typically not appropriate for this high degree of censoring and thus a 321 binomial method was used. Equations 7 and 8 were used for showering, garden hose use, car 322 washing, and toilet flushing, accounting for the volume of aerosols of various size diameters that are 323 large enough to hold LP or MAC but small enough to deposit at the alveoli (1µm < diameter < 10µm). 324

325
$$Dose_{LP,j} = \frac{1}{R} C_{RHRW,LP} \frac{1}{n_s} P_{contam} Bt \sum_{i=1}^n C_{aer,i} V_{aer,i} DE_i$$
(7)

326

327

$$Dose_{MAC,j} = \frac{1}{R} C_{RHRW,MAC} Bt \sum_{i=1}^{n} C_{aer,i} V_{aer,i} DE_i$$
(8)

328

329 Where n_s= number of samples, P_{contam}= binomial probability of contamination with number of samples 330 n_s and probability of contamination p, $C_{aer,i}$ = the concentration of aerosols of diameter i where i =1:10, $V_{aer,i}$ = the volume of each aerosol size bin *i* calculated as $V = (4/3)\pi r^3$, B = breathing rate (m³/min), *t* 331 = exposure duration (min); and DE = alveolar deposition efficiency of size *i* diameter aerosols. 332 The aerosol size distributions for aerosols of diameters 1-10 µm from toilet flushing, showering, 333 and hose use are provided by O'Toole et al. (O'Toole et al., 2008b, O'Toole et al., 2009) and are 334 335 summarized in Table 2. For showering, a conventional showerhead operating at a water temperature 336 of 42°C was chosen. For toilet flushing, a full 9 L flush 420 mm above the toilet was chosen; this 337 category only observed aerosols in one size bin (median 2.5 µm diameter). For gardening, hose use for gardening purposes is assumed to use a "conventional" nozzle on a "spray" setting while car 338 339 washing would be represented by hosing with a high pressure "water efficient device" on a "jet" 340 setting. Experiments with the largest generation of aerosols in the 1-10 µm diameter range were 341 chosen.

342 2.3.2. Pool top-up

Partitioning coefficients were used to model swimming in a pool that has been partially filled with 343 344 RHRW using equations 9 and 10.

345

346

$$P_{LP,pool} = \frac{1}{R} C_{RHRW,LP} \frac{1}{134} P_{contam} P_{LP} B t_{pool} D$$
(9)

D R +

$$D_{MAC,pool} = \frac{1}{R} C_{RHRW,MAC} P_{MAC} B t_{pool} D$$
(10)

 $-\frac{1}{2}C_{-1}$

347

Where P = water to air partitioning coefficient (L per m³), B = breathing rate (m³ per min), and t = 348 exposure duration (min). Dilution D was assessed for the same conditions as the ingestion model. 349 350 The partitioning coefficient for LP in an indoor warm therapy pool was calculated to range from 2.2 x 10^{-8} to 1.1×10^{-5} L per m³ in a previous study (Hines et al., 2014). A partitioning coefficient for MAC 351 352 was calculated in the current study using ratios of median concentrations of nontuberculous 353 mycobacteria in therapy pool water and air reported by Glazer et al., (2007) for sites where NTM was recovered from both air and water ranging from 1.0×10^{-4} to 5.3×10^{-3} L per m³. The number of 354 minutes of exposure time per swim was derived from a published study (Schets et al., 2011) by 355 356 combining parameters for males, females, and children using the same method as for swimming

exposure frequency (section 2.2.6). Bathing for hygienic purposes was not considered for inhalation(general population) exposures.

359 2.3.3. Opportunistic pathogen concentrations in RHRW

The concentrations of LP, *M. avium*, and *M. intracellulare* ranged up to 9.8×10^3 gene copies per L, 360 361 1.1×10^5 gene copies per L, and 6.8×10^5 gene copies per L, respectively (Hamilton et al., 2016). *M*. 362 intracellulare is thought to only reside in soil, while M. avium or M. chimaera are found in water 363 (Wallace et al., 2013). The primer set used in Hamilton et al., 2016 for *M. intracellulare* (Chern et al., 364 2015) identifies both *M. intracellulare* and *M. chimaera*. As *M. intracellulare* is not expected to be present in water, we assume all M. intracellulare enumerated were actually M. chimaera. 365 366 Furthermore, because of the lack of a dose response model specific to *M. chimaera*, we assumed that 367 the dose response relationships in Hamilton et al., 2017 can be applied to the total dose of MAC (M. 368 avium and M. chimaera). This presents a challenge for adding these on a sample by sample basis 369 where, for example, the concentration of *M. avium* was positive (above the lower limit of detection, 370 LLOD) and above the lower limit of quantification (LLOQ), but the concentration of *M. intracellulare* was positive (above the LLOD) but above the LLOQ. To address this issue, separate interval-371 372 censored lognormal distributions were fitted to each dataset (M. avium and M. intracellulare) using the 373 package fitdistrplus in R, and the simulated distributions were added within the Monte Carlo 374 simulation model to obtain the total MAC count. Therefore, $C_{RHRW, MAC} = C_{RHRW, MA} + C_{RHRW, MI}$ (Table 1). 375 It is noted that in the original study, there was no significant differences in pathogen concentrations by 376 sampling cluster (Brisbane vs. Currumbin Ecovillage) (Hamilton et al. 2016). The primer sets for M. 377 avium and M. intracellulare both target the 16S gene, which occurs as a single copy (Chern et al. 378 2014). Similarly, L. pneumophila primer sets targeted the mip gene which is a single copy gene 379 (Engleberg et al. 1989). It was therefore assumed that one gene copy was equivalent to one viable, 380 infectious microorganism for *M. avium*, *M. intracellulare*, and LP.

381 2.4. Dose response

Exponential and Beta-Poisson dose response models (Haas et al. 1999) are stated in equations 11 and 12, respectively, with modifications as described below. Exponential dose response model parameters for LP infection are provided in Table 3 (Armstrong and Haas, 2007). Exponential and Beta-Poisson dose response models for MAC pulmonary infection, disseminated infection, and cervical lymphadenitis were used (Jorgensen et al. 1977 model (1) for cervical lymphadenitis and the 387 Yangco et al. 1989 model for disseminated infection were chosen and are described in detail in 388 relation to these endpoints elsewhere (Hamilton et al., 2017). Of the three available cervical 389 lymphadenitis dose-response models (Hamilton et al. 2017), the model with a lymph node lesions 390 endpoint (instead of infection) was selected as this was likely to describe a more severe infection 391 representative of cervical lymphadenitis in children. A conversion factor (C) for pulmonary infection of 392 500 is applied to convert the model from the intravenous route to the inhalation route as per Hamilton 393 et al., 2017, by dividing the daily dose by 500 within the inhalation models only. For all other models, 394 C = 1.

$$P_{inf,daily} = 1 - e^{-rd/C}$$
(11)

(12)

 $P_{inf,daily} = 1 - (1 + \frac{d/c}{\beta})^{-\alpha}$

397 Where $P_{inf,daily}$ = daily probability of infection, d = daily dose, and r, α , and β are parameters of the 398 respective dose response models.

For the special case of lymphadenitis in children, disease occurs only in children 1-12 years (Lai et al. 1984) of age and the majority of cases are reported in children 1-5 (Inderlied et al. 1993, Haverkamp et al. 2004, Wolinsky 1995, Lincoln et al, 1972, Hazra et al. 1999, Tebruegge et al. 2016). To account for variability in age groups and difference in susceptibilities among the child population at risk, the daily probability of infection is calculated for children according to equation 13:

404

$$P_{inf,daily,child,activity} = 1 - \prod_{i=1}^{l} \left(1 - P_{inf,daily,i}P_{attr_{i}}\right)$$
(13)

406

Where $P_{inf, daily, child, activity}$ = daily probability of MAC cervical lymphadenitis due to a given ingestion activity (swimming, toilet flushing, etc.) in a child from 1-12 years of age; $P_{inf, daily, i}$ = calculated daily probability of lymphadenitis for children in age bracket *i* where *i* = 1-2, 3-5, or 6-12), P_{attr} = Portion of total age 1-12 MAC-attributable lymphadenitis cases in age bracket *i*. Lai et al. (1984) report that 87% of total diagnosed mycobacterial cervical lymphadenitis cases occurred before age 12. Of the total cases occurring from ages 1-12, 60% occurred between ages 1-3, 27% occurred from ages 3-5, and 13% occurred from ages 5-12; these values are used as $P_{attr, i}$.

For children ages 1-2, the exposure pathways considered are drinking water, eating lettuce,
bathing, and swimming in pools topped-up with rainwater. For children ages 3-5, the exposure

pathways considered are drinking, eating lettuce, bathing (instead of showering), swimming in pools
topped-up with rainwater, and toilet flushing. For ages 6-12, exposure scenarios considered were
considered were drinking, eating lettuce, showering, swimming in pools topped-up with rainwater, and
toilet flushing.

420

421 2.5. Risk characterization

422 Annual risk was calculated as per Equation 14.

423

 $P_{inf,ann} = 1 - \prod_{1}^{nf_j} (1 - P_{inf,daily})$

425

Where *n* is the yearly frequency and *f* is the daily frequency of the activity *j*. Frequency $f_j = 1$ unless otherwise stated in Table 2. A sensitivity analysis was conducted to identify variables contributing to uncertainty using 100,000 Monte Carlo iterations. All computations were performed in R (www.rproject.org) and using the mc2d package (Pouillot et al., 2010). Random sampling of daily risks with replacement was conducted as per the preferred method for annualizing probability of infection using 100,000 iterations (Karavarsamis and Hamilton, 2010). In addition to annual risks for each exposure scenario, total annual infection risks for each

population were calculated according to equation 15. A similar approach has been used to pool risks
from multiple pathogens by previous QMRA studies (de Man et al. 2014b, Soller et al. 2016, Ahmed
et al. 2010).

436

$$P_{inf,ann,total,s,e,p,c} = 1 - \prod_{1}^{J} (1 - X_{a_j} P_{inf,ann,a_j})$$
(15)

437

Where $P_{inf,ann,total,s,e,p,l}$ = the total annual risk incurred from each scenario *s* where *s* = inhalation or ingestion annual risk from *j* activities a_1 , a_2 , $a_3..a_j$ where a_j = showering, drinking, etc., and $X_{a,j}$ = portion of sampling cluster location *c* that uses rainwater for each activity (Supplemental Table 1). For child bathing in the ingestion scenario, because exposure volumes were not available, it was assumed that the same volume is ingested during indoor bathing for hygienic purposes in a bathtub as during swimming in an outdoor pool (Schets et al. 2011). Total risk is calculated for health endpoint *e* = pulmonary infection, cervical lymphadenitis, or disseminated infection and *p* = healthy 445 populations, children, or vulnerable/immune-compromised populations that are relevant for each of 446 the previous endpoints, respectively. Total risks for each scenario are specific to each rainwater-using 447 population group p, and are therefore not extrapolated to a general population level that includes non-448 rainwater users. For the pulmonary infection endpoint, LP and MAC risks are included in the same 449 equation to compute total pulmonary disease risks, assuming the infection risks from these organisms 450 are additive and that infection with one pathogen would not preclude infection with the other. 451 The Spearman rank correlation coefficient was used to identify the most important predictive 452 factors of annual infection or clinical severity infection risk, were 0 is no influence and -1 or +1 when 453 the output is wholly dependent on that input. The model inputs were ranked based on their correlation

454 coefficient with the output variable, annual risk.

455

456 **3. Results**

457 3.1 Ingestion

Annual risks for each ingestion scenario are shown in Fig. 2 and compared to a hypothetical 1 × 10⁻⁴ 458 drinking water annual infection risk benchmark (Regli et al., 1991). Consumption of rainwater 459 460 presented the highest risk for both cervical lymphadenitis in children and disseminated infection for 461 immune-compromised adults. The use of drinking water POU filters reduced total risks by approximately 90% (<1 log), but did not bring drinking water risks below 10⁻⁴. For children, bathing 462 463 (instead of showering) in untreated, undiluted rainwater also resulted in high annual risks above 10⁻⁴. In all drinking water scenarios, the median annual risk was > 1×10^{-4} . For child non-recreational 464 465 exposures, consumption of lettuce irrigated with RHRW, showering, and toilet flushing had median annual lymphadenitis risks below the benchmark, however the 95[%] confidence limit for toilet flushing 466 467 exceeded the benchmark. For immune-compromised non-recreational scenarios, only car washing 468 and clothes- washing had median annual disseminated infection risks below the benchmark and the 469 95% confidence limit for car washing exceeded this value. There is no risk benchmark for swimming 470 in pools, although there are recreational standards for microbiological risks associated with freshwater 471 and marine beaches of 8 and 19 cases of highly credible gastrointestinal illness (HCGI) per 1,000 472 recreators in fresh and marine waters, respectively (US EPA, 2011b). Eight cases of HCGI per 1,000 473 is equivalent to 36 cases of gastrointestinal illness per 1,000 recreators based on a more 474 encompassing definition of gastrointestinal illness used in the USEPA National Epidemiological and

475 Environmental assessment of Recreational Water (NEEAR) studies (USEPA, 2011b). These 476 definitions are not particularly compatible with the health endpoints used in this analysis, and are 477 therefore not used for comparison here. Generally, swimming pool risks were higher for 478 vulnerable/immune-deficient populations than for children. For child exposures, swimming pool water 479 would need to be diluted to 50% RHRW: 50% sterile water in order to have a median risk below or equal to 1×10^{-4} for children swimming at an average rate (n= 32 swims per year) and 10% RHRW: 480 sterile water for children with an upper-bound exposure based on an upper bound 95th percentile 481 estimate of 122 swims per year. Of the dilution scenarios tested (10%, 50%, and 90% RHRW: sterile 482 water), the median annual pool risks for immune-compromised individuals were never below 10⁻⁴. If 483 using a less stringent recreational water-based risk comparison point of approximately 10⁻² on an 484 485 annual basis, a dilution of 10% RHRW: sterile water would be needed to achieve median annual risks 486 below this level for disseminated infection. All child median annual risks were below this level. 487 A sensitivity analysis for ingestion risk scenarios is shown in Table 6. The most influential 488 predictor of variability in annual risk for all scenarios was the concentration of MAC in RHRW (CMA. 489 C_{M}) (Spearman rank correlation coefficients ranging from 0.16-0.19 and 0.70- 0.90 for MA and MI, 490 respectively). However, as C_{RHRW} was calculated by adding the concentration of *M. avium* and *M.* 491 intracellulare in RHRW, the sensitivity analysis determined that the concentration of M. intracellulare 492 was more influential as this is more commonly present in RHRW and in higher concentrations than M. 493 avium on average. The dose response parameter r and volume of water ingested (V_{ing}) or retained by lettuce (V_R) were the second and third most influential parameters, respectively. 494

495 3.2 Inhalation

496 Annual risks for each inhalation scenario are shown in Fig. 3. The median annual LP risks for 497 showering, and garden hosing exceeded a 1×10^{-4} benchmark. Risks for toilet flushing and car 498 washing were below the benchmark. Risks were highest for showering and pool top-up. For MAC, all 499 95% confidence intervals were below the benchmark value. Annual pool swimming risks were above 500 10^{-4} but below 10^{-2} .

A sensitivity analysis for inhalation risk scenarios is shown in Table 6. The most influential predictor of variability in daily risk for toilet flushing, showering, garden hosing, or car washing was the concentration of LP and/or MAC in RHRW (C_{RHRW} , C_{Ml} , or C_{MA}) (Spearman rank correlation coefficients ranging from 0.14- 0.88). For LP toilet flushing scenario only, the concentration of aerosols (C_{aer}) was

- the most important predictor ($\rho = 0.80$) while for MAC toilet flushing, C_{aer} was the second most
- 506 important predictor (ρ = 0.58). For LP scenarios, the probability of contamination (P_{contam}) was also an
- 507 important factor (Spearman rank correlation coefficients ranging from 0.26 to 0.47). For all pool
- 508 scenarios, the partitioning coefficient (*P*) was either the first or second-most important predictor
- 509 (Spearman rank correlation coefficients ranging from 0.29 to 0.50).
- 510 3.3 Total annual risks

511 In order to quantify the risks for each relevant population/ exposure route, a distribution of total annual 512 risks was computed for each scenario. For cervical lymphadenitis in children and disseminated 513 infection in immune-compromised populations via a combination of all oral ingestion exposure routes 514 (Fig. 4), risks for immune-compromised populations were higher than for children, and higher for the 515 Currumbin Ecovillage study population than the Brisbane study population based on survey 516 responses. Total annual risks were not substantially impacted by the dilution of RHRW used in pools 517 with sterile water or the number of swims per year The use of drinking water POU filters reduced total risks by approximately 90% (<1 log). All median total risks were above a 1×10^{-4} benchmark. 518 519 Total inhalation risks from all aerosol-generating activities reported in the survey for each 520 sampling cluster are summarized in Fig. 5. Median total annual LP pulmonary infection risks were approximately 4-5 orders of magnitude higher than MAC pulmonary infection risks for all scenarios, 521 and all 95% confidence intervals for LP total risks were above the 1 × 10⁻⁴ benchmark while all those 522 523 for MAC were below this value. Therefore, total pulmonary infection risks (LP + MAC) were driven by 524 LP risks rather than MAC risks. Total annual risks for LP were slightly higher for the Brisbane 525 compared to the Currumbin Ecovillage, while total annual MAC risks were lower for Currumbin than 526 for Brisbane. The number of swims per year and pool dilution did not have a substantial impact on 527 differences in risk between scenarios.

528

529 4. Discussion

530 The current study is the first investigation to assess MAC risks for human-relevant, non-MAP 531 species of MAC in an aquatic environment. For MAC, the drinking water route of exposure presents 532 the greatest risks for susceptible populations including children and the immune-compromised. For 533 these populations, the simulated risks do not support the conclusion that it would be appropriate to 534 drink rainwater or use it for bathing children without treatment. The use of a filter reduced risks but did not reduce them to below benchmark levels; additionally, opportunistic pathogens such as MAC can
colonize POU filtration systems (Rodgers et al., 1999), limiting their utility for mitigating drinking water
risks in these circumstances.

538 For swimming pool use, risks can be lowered through dilution with treated water. Although there is 539 no risk benchmark for opportunistic pathogens in pools, this work indicates that a dilution to 90% 540 rainwater would bring median risks below a recreational benchmark. However, the risk simulations 541 conducted do not indicate that it would be feasible to reduce annual risks for immune-compromised populations to below 10⁻⁴ but for children, a median annual risk would be below this level for 50% 542 dilution RHRW: sterile water. A 10⁻⁴ risk level may be more appropriate for comparison in order to 543 544 reduce risks in this case. However, this dilution factor for children can be lessened depending on the 545 recreational water quality criteria used for comparison. Legionella pneumophila risks were above the 546 drinking water benchmark for all scenarios except toilet flushing and car washing, and were greater 547 than MAC risks in all cases. Showering and swimming pool use presented the highest risks for 548 pulmonary infection.

Both inhalation and ingestion simulations indicate that for non- immune-compromised populations, 549 550 toilet flushing is one of the lowest-risk scenarios and is likely to be an appropriate use for rainwater. 551 Similarly, clothes washing and car washing represented the two other lower-risk scenarios. Clothes 552 washing, garden hose use, and car washing scenarios were not considered for children; however 553 some children may perform these types of chores, although it is likely to be a small percentage of 554 children and there is likely to be considerable variability in these behaviors (Klein et al. 2009). For 555 children, consumption of lettuce was also below the benchmark risk. All individual MAC pulmonary 556 infection inhalation risks for healthy populations were below the drinking water benchmark. This could 557 be affected by the fact that a conversion factor of 500 was used to convert the intravenous to 558 inhalation dose response route (Hamilton et al., 2017). Due to the lack of dose response models for 559 pulmonary infections in susceptible populations, further investigation of potential dose response 560 models for this purpose is recommended. When considering "total" risks for populations performing all 561 the possible activities using rainwater, the risk simulations indicated that total risks could be high and in no case was below 10⁻⁴. This supports that using RHRW as a sole source of water could incur 562 563 potential risks and additional assessment of combinations of activity patterns yielding risks within 564 certain ranges would be beneficial for decision-making purposes.

565 An important factor in this work was the regional preference for various uses of RHRW. When 566 examining differences between two sampling clusters in Brisbane and the Currumbin Ecovillage, 567 ingestion risks and inhalation risks for total inhalation pathogen load (LP and MAC) were higher for 568 the Currumbin Ecovillage compared to Brisbane. This is likely due to the higher degree of RHRW 569 application for potable uses in Currumbin compared to Brisbane. When examining total inhalation 570 risks from all uses (showering, garden hose, car washing, etc.) and considering L. pneumophila and 571 MAC risks separately, Currumbin risks for LP were higher than Brisbane but MAC risks were lower 572 than Brisbane. This may be due to higher RHRW use for pools and showers in Brisbane than Currumbin. MAC pool inhalation risks were computed using a partitioning coefficient, which is 573 574 reported to be higher for MAC than for LP. L. pneumophila shower risks were the highest compared to 575 other scenarios, indicating that showering may have had a greater influence on total annual 576 pulmonary risk for Currumbin compared to the other scenarios.

For previous studies focusing on LP risks in RHRW with LP concentrations of 60 to 170 gene copies (assumed equivalent to cells) per L rainwater, between 3.0×10^{-2} and 8.6×10^{-2} LP infections per 10,000 exposed people per shower exposure and 1.8×10^{-2} - 5.1×10^{-2} infections per 10,000 exposed people per hosing exposure were expected (Ahmed et al., 2010). This would be between an annual probability of infection of approximately 1.09×10^{-3} and 3.13×10^{-3} and is comparable with the risks calculated in the current study. A case of Legionnaires disease has also been previously linked to the usage of a garden hose (Piso et al., 2007).

584 Another study of rainwater used as a source of water for splash parks estimated a mean Legionella infection risk of 9.3×10^{-5} for a 3.5 minute splash park exposure for children and 1.1×10^{-4} 585 for adults, but that for a 2 h exposure this risk would be approximately 2.8×10^{-3} (de Man et al., 586 587 2014a). Although the pool exposure model is different than that used in the current study, de Man et al. work supports that pool risks can be potentially high compared to other exposure routes. The 588 589 current QMRA did not consider ingestion of mouthfuls of water as in de Man et al. models due to the 590 difference between active splash parks and private pools. However, exposures to pool water could 591 potentially be higher in some circumstances if mouthfuls of water are considered. Private household 592 pool or whirlpool use has indeed been linked to Legionnaires disease cases (Euser et al., 2010). It is 593 noted in the current QMRA that although dilution of RHRW with sterile water was considered, RHRW 594 users may dilute rainwater with treated tap-water which may not be free of LP and MAC but is likely to contain fewer pathogens than RHRW (Whiley et al., 2014). Additionally, some chlorine residual
present in municipal drinking water could contribute to pathogen removal. However, LP (Cooper and
Hanlon, 2010), and especially MAC (Falkinham 3rd, 2003; Steed and Falkinham 3rd, 2006; Taylor et
al., 2000) are resistant to disinfection including chlorination, and would not be likely to decline
significantly at low levels of chlorine residual. Chlorine treatment in pools by owners could contribute
to pathogen removal and could be considered in a more detailed assessment of opportunistic
pathogens in pools.

Similarly to the previous QMRAs for RHRW (Ahmed et al., 2010; de Man et al., 2014a), in the absence of information regarding the relationship between total gene copies and the viability of pathogens in rainwater tanks, one gene copy was considered equivalent to one viable, infectious pathogen. This assumption could potentially lead to an overestimation of risks; however, culturebased assessments of pathogens can neglect to quantify viable but non-culturable (VBNC) microorganisms, and therefore underestimate risks. If a 1% or 10% viability assumption is made, total annual risks would be 1 to 2 orders of magnitude lower, respectively.

Finally, regarding the clothes washing scenario, MAC can be present in fecal material (Yajko et
al., 1993) that can be introduced to the laundry cycle through soiled clothing and provide additional
pathogen loading. A more in-depth QMRA model for MAC risks due to clothes washing might
consider transfer of fecal-associated MAC to water and hand-to-mouth contact after clothes
laundering and impact of any disinfectants used (Callewaert et al., 2015; Gerba and Kennedy, 2007,
Gibson et al., 1999; Lopez et al., 2013).

615 Regarding the management of RHRW risks, toilet lids can be closed prior to flushing to prevent 616 the spread of mists. Similarly, less aerosol-efficient showerheads can be installed in showers for 617 which rainwater is used as a source. For pool-users, pool filters can be installed and rainwater can be 618 diluted with treated water in order to potentially lower risks. In this study, only under-sink POU filters 619 were considered for mitigating drinking water risks, however, other methods such as solar-disinfection 620 have been explored for use with rainwater tanks which could provide higher log removals (Reyneke et 621 al., 2016). Treatment systems designed to treat all water entering the home from a rainwater tank as 622 opposed to only water obtained via a kitchen sink tap would be beneficial for reducing risks. More exploration of in situ field performance of various rainwater treatment systems over time is warranted 623 624 as a means for approaching pathogen risk mitigation for RHRW.

5. Conclusions

627	٠	Regional use preferences for roof-harvested rainwater had an impact on the rank order of
628		ingestion and inhalation risks for opportunistic pathogens
629	•	Based on the results of the Mycobacterium avium quantitative microbial risk assessment
630		ingestion scenarios, harvested rainwater is not recommended for drinking in child or immune-
631		compromised populations
632	•	For healthy populations, showering and garden hosing are not recommended for harvested
633		rainwater
634	•	Compared to Mycobacterium avium infection risks, potential Legionella risks should drive risk
635		mitigation strategies for the inhalation route of exposure
636		
637	Ackn	owledgements
638	This wo	ork was supported by a Fulbright-CSIRO Postgraduate scholarship sponsored by the
639	Austral	ian-American Fulbright Commission. The authors are grateful to rainwater study participants
640	for prov	viding survey information used in this work and for CSIRO staff Dr. Jatinder Sidhu, Leonie
641	Hodge	rs, Andrew Palmer, Kylie Smith, and Pradip Gyawali for their contributions to the original
642	pathog	en quantification study used in this risk assessment.
643		
644		
645		
646		
647		X ·

653

- 654
- 655
- 656
- 657
- 658
- ~ ~
- 659
- 660
- 661

662 **References**

663 ABS, 1998. 4359.0 1995 How Australians measure up. Canberra.

664 ABS, 2010. Environmental issues: water use and conservation, Australian Bureau of Statistics, Canberra, 665 Australia.

666 ABS, 2016. 3235.0 – Population by age and sex, regions of Australia, 2015. Available at

http://www.abs.gov.au/ausstats/abs@.nsf/Latestproducts/3235.0Main%20Features252015?opendocu
 ment&tabname=Summary&prodno=3235.0&issue=2015&num=&view=.

ment&tabname=Summary&prodno=3235.0&issue=2015&num=&view=.
Ahammed, M.M., Meera, V., 2010. Metal oxide/hydroxide-coated dual-media filter for simultaneous
removal of bacteria and heavy metals from natural waters. J. Hazarad Mater. 181 (1), 788-793.
Ahmed, W., Brandes, H., Gyawali, P., Sidhu, J.P.S., Toze, S., 2014. Opportunistic pathogens in roofcaptured rainwater samples, determined using quantitative PCR. Water Res. 53, 361-369.
Ahmed, W., Hamilton, K., Vieritz, A., Powell, D., Goonetilleke, A., Hamilton, M., Gardner, T., 2016.

674 Microbial risk from source-separated urine used as liquid fertilizer in sub-tropical Australia. Microbial Risk 675 Anal. (on-line early).

676 Ahmed, W., Hodgers, L., Masters, N., Sidhu, J.P., Katouli, M., Toze, S., 2011. Occurrence of intestinal
and extraintestinal virulence genes in *Escherichia coli* isolates from rainwater tanks in Southeast
Queensland, Australia. Appl. Environ. Microbiol. 77 (20), 7394-7400.

679 Ahmed, W., Vieritz, A., Goonetilleke, A., Gardner, T., 2010. Health risk from the use of roof-harvested
rainwater in Southeast Queensland, Australia, as potable or nonpotable water, determined using
quantitative microbial risk assessment. Appl. Environ. Microbiol. 76 (22), 7382-7391.

682 Armstrong, T., Haas, C.N., 2007. A quantitative microbial risk assessment model for Legionnaires' 683 Disease: animal model selection and dose-response modeling. Risk Anal. 27 (6), 1581-1596.

684 Australian Government Bureau of Meteorology., 2016. Climate Data Online.

685 Australian Government Department of Health., 2016. Notifications of a selected disease by month and 686 year.

687 Barker, S.F., O'Toole, J., Sinclair, M.I., Leder, K., Malawaraarachchi, M., Hamilton, A.J., 2013. A probabilistic model of norovirus disease burden associated with greywater irrigation of homeproduced lettuce in Melbourne, Australia. Water Res. 47 (3), 1421-1432.

690 Bloom, D.A., Seeley, W.W., Ritchey, M.L., McGuire, E.J., 1993. Toilet habits and continence in children:

an opportunity sampling in search of normal parameters. J. Urol. 149 (5), 1087-1090.

692 Breuninger, K.J., Weir, M.H., 2015. Development of an interspecies nested dose-response model for 693 *Mycobacterium avium* subspecies *paratuberculosis*. Risk Anal. 35 (8), 1479-1487.

694Brodribb, R., Webster, P., Farrel, D., 1995. Recurrent *Campylobacter fetus* subspecies bacteraemia in a 695 febrile neutropaenicpatient linked to tank water. Commun. Dis. Intel. 19 (13), 312-313.

696 Callewaert, C., Van Nevel, S., Kerckhof, F.-M., Granitsiotis, M.S., Boon, N., 2015. Bacterial exchange in 697 household washing machines. Front. Microbiol. 6, 1381.

698 Chern, E.C., King, D., Haugland, R., Pfaller, S., 2015. Evaluation of quantitative polymerase chain 699 reaction assays targeting *Mycobacterium avium*, *M. intracellulare*, and *M. avium* subspecies

paratuberculosis in drinking water biofilms. J. Water Health. 13 (1), 131-139.

701 Cook, K., Flis, S., Ballard, C., 2013. Sensitivity of *Mycobacterium avium* subsp paratuberculosis, 702 Escherichia coli and Salmonella enterica serotype Typhimurium to low pH, high organic acids and 703 ensiling. J. Appl. Microbiol. 115 (2), 334-345. 704 Cooper, I., Hanlon, G., 2010. Resistance of Legionella pneumophila serotype 1 biofilms to chlorine-based 705 disinfection. J. Hosp. Infect. 74 (2), 152-159. 706 Crabtree, K.R., Ruskin, R.H., Shaw, S.B., Rose, J.B., 1996. The detection of Cryptosporidium oocysts 707 and Giardia cysts in cistern water in the US Virgin Islands. Water Res. 30 (1), 208-216. 708 CSIRO, 2008. 2007 Australian national children's nutrition and physical activity survey: main findings. Australian Government Department of Health and Ageing, Australian Food and Grocery Council, 709 Australian Government Department of Agriculture, Fisheries and Forestry. 710 711 de Man, H., Bouwknegt, M., van Heijnsbergen, E., Leenen, E.J., van Knapen, F., de Roda Husman, A.M., 712 2014a. Health risk assessment for splash parks that use rainwater as source water. Water Res. 54, 713 254-261. 714 de Man. H., van den Berg. H., Leenen, E., Schiiven, J., Schets, F., van der Vliet, J., van Knapen, F., de Roda Husman, A., 2014b. Quantitative assessment of infection risk from exposure to waterborne 715 pathogens in urban floodwater. Water Res. 48, 90-99. 716 717 Dicaprio, E., Ma, Y., Purgianto, A., Hughes, J., Li, J., 2012. Internalization and dissemination of human norovirus and animal caliciviruses in hydroponically grown romaine lettuce. Appl. Environ. Microbiol. 718 719 78 (17), 6143-6152. 720 Diederen, B., 2008. Legionella spp. and Legionnaires' disease. J. Infect. 56 (1), 1-12. 721 Dobrowsky, P., Carstens, M., De Villiers, J., Cloete, T., Khan, W., 2015a. Efficiency of a closed-coupled solar pasteurization system in treating roof harvested rainwater. Sci. Total. Environ. 536, 206-214. 722 723 Dobrowsky, P., Lombard, M., Cloete, W., Saayman, M., Cloete, T., Carstens, M., Khan, S., Khan, W., 2015b. Efficiency of microfiltration systems for the removal of bacterial and viral contaminants from 724 725 surface and rainwater. Water Air Soil Pollut. 226 (3), 1-14. 726 Dufour, A.P., Evans, O., Behymer, T.D., Cantu, R., 2006. Water ingestion during swimming activities in a pool: a pilot study. J. Wat. Health. 4 (4), 425-430. 727 728 Egodawatta, P., Thomas, E., Goonetilleke, A., 2009. Understanding the physical processes of pollutant build-up and wash-off on roof surfaces. Sci. Total. Environ. 407 (6), 1834-1841. 729 730 Ellis, K., 1993. Legionellosis: A concise review. Water Environment J. 7 (4), 418-430. 731 Engelberg, N.C., Carter, C., Weber, D.R., Cianciotto, N.P., Eisenstein, B.I., 1989. DNA sequence of mip, 732 a Legionella pneumophila gene associated with macrophage infectivity. Infect. Immun. 57 (4), 1263-733 1270. 734 Euser, S.M., Pelgrim, M., Den Boer, J.W., 2010. Legionnaires' disease and Pontiac fever after using a 735 private outdoor whirlpool spa. Scand. J. Infect. Dis. 42 (11-12), 910-916. 736 Falkinham 3rd, J., Hilborn, E.D., Arduino, M.J., Pruden, A., Edwards, M.A., 2015. Epidemiology and 737 ecology of opportunistic premise plumbing pathogens: Legionella pneumophila, Mycobacterium avium, and Pseudomonas aeruginosa. Environ. Health Perspect. 123 (8), 749-758. 738 739 Falkinham 3rd, J.O., 1996. Epidemiology of infection by nontuberculous mycobacteria. Clin. Microbiol. 740 Rev. 9 (2), 177-215. 741 Falkinham 3rd, J.O. (2003) Factors influencing the chlorine susceptibility of *Mycobacterium avium*, Mycobacterium intracellulare, and Mycobacterium scrofulaceum. Applied and Environmental 742 Microbiology 69(9), 5685-5689. 743 744 Falkinham 3rd, J.O., 2013. Ecology of nontuberculous Mycobacteria - where do human infections come from? Semin, Respir. Crit. Care Med. 34 (1), 95-102. 745 746 Falkinham 3rd, J.O., Pruden, A., Edwards, M., 2015. Opportunistic premise plumbing pathogens: Increasingly important pathogens in drinking water. Pathogens. 4 (2), 373-386. 747 748 Fewtrell, L., Kay, D., 2007. Quantitative microbial risk assessment with respect to Campylobacter spp. in 749 toilets flushed with harvested rainwater. Water Environ. J. 21 (4), 275-280. 750 Field, S.K., Fisher, D., Cowie, R.L., 2004. Mycobacterium avium complex pulmonary disease in patients without HIV infection. Chest 126 (2), 566-581. 751 752 Franklin, L.J., Fielding, J.E., Gregory, J., Gullan, L., Lightfoot, D., Poznanski, S.Y., Vally, H., 2009. An 753 outbreak of Salmonella Typhimurium 9 at a school camp linked to contamination of rainwater tanks. 754 Epidemiol. Infect. 137 (3), 434-440. 755 Freeman, A.F., Olivier, K.N., Rubio, T.T., Bartlett, G., Ochi, J.W., Claypool, R.J., Ding, L., Kuhns, D.B., Holland, S.M., 2009. Intrathoracic nontuberculous mycobacterial infections in otherwise healthy 756 757 children. Pediatr. Pulmonol. 44 (11), 1051-1056. 758 Gerba, C.P., Kennedy, D., 2007. Enteric virus survival during household laundering and impact of 759 disinfection with sodium hypochlorite. Appl. Environ. Microbiol. 73 (14), 4425-4428.

760 Gibson, L.L., Rose, J.B., Haas, C.N., 1999. Use of quantitative microbial risk assessment for evaluation of the benefits of laundry sanitation. Am. J. Infect. Control. 27 (6), S34-S39.

762 Glazer, C.S., Martyny, J.W., Lee, B., Sanchez, T.L., Sells, T.M., Newman, L.S., Murphy, J., Heifets, L.,
Rose, C.S., 2007. Nontuberculous mycobacteria in aerosol droplets and bulk water samples from
therapy pools and hot tubs. J. Occup. Environ. Hyg. 4 (11), 831-840.

765 Griffith, D.E., Aksamit, T., Brown-Elliott, B.A., Catanzaro, A., Daley, C., Gordin, F., 2007. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial

767 diseases. Am. J. Resp. Crit. Care Med. 175 (4), 367-416.

768 Haas, C.N., Rose, J.B., Gerba, C.P., 1999. Quantitative microbial risk assessment, John Wiley & Sons.

769 Hamilton, K., Ahmed, W., Palmer, A., Sidhu, J., Hodgers, L., Toze, S., Haas, C., 2016. Public health
implications of *Acanthamoeba* and multiple potential opportunistic pathogens in roof-harvested
rainwater tanks. Environ. Res. 150, 320-327.

772 Hamilton, K.A., Weir, M.H., Haas, C.N., 2017. Dose response models and a quantitative microbial risk
assessment framework for the *Mycobacterium avium* complex that account for recent developments
in molecular biology, taxonomy, and epidemiology. Water Res. 109, 310-326.

775 Haverkamp, M.H., Arend, S.M., Lindeboom, J.A., Hartwig, N.G., van Dissel, J.T., 2004. Nontuberculous
 mycobacterial infection in children: a 2-year prospective surveillance study in the Netherlands. Clin.
 Infect. Dis. 39 (4), 450-456.

778 Hazra, R., Robson, C.D., Perez-Atayde, A.R., Husson, R.N., 1999. Lymphadenitis due to nontuberculous

779 mycobacteria in children: presentation and response to therapy. Clin. Infect. Dis. 28 (1), 123-129.

780 Heyder, J., Gebhart, J., Rudolf, G., Schiller, C.F., Stahlhofen, W., 1986. Deposition of particles in the 781 human respiratory tract in the size range 0.005-15 μm. J. Aerosol Sci. 17 (5), 811-825.

782 Hines, S.A., Chappie, D.J., Lordo, R.A., Miller, B.D., Janke, R.J., Lindquist, H.A., Fox, K.R., Ernst, H.S.,
783 Taft, S.C., 2014. Assessment of relative potential for *Legionella* species or surrogates inhalation
784 exposure from common water uses. Water Res. 56, 203-213.

785 Hirneisen, K.A., Sharma, M., Kniel, K.E., 2012. Human enteric pathogen internalization by root uptake into food crops. Foodborne Pathog. Dis. 9 (5), 396-405.

787 Holvoet, K., De Keuckelaere, A., Sampers, I., Van Haute, S., Stals, A., Uyttendaele, M., 2014.

788 Quantitative study of cross-contamination with *Escherichia coli*, *E. coli* O157, MS2 phage and murine 789 norovirus in a simulated fresh-cut lettuce wash process. Food Control. 37, 218-227.

790 Inderlied, C.B., Kemper, C.A., Bermudez, L.E., 1993. The *Mycobacterium avium* complex. Clin. Microbiol. 791 Rev. 6 (3), 266-310.

792 Jordan, F.S., Seaman, R., Riley, J.J., Yoklic, M.R., 2008. Effective removal of microbial contamination

from harvested rainwater using a simple point of use filtration and UV-disinfection device. Urban
 Water J. 5 (3), 209-218.

795 Kaevska, M., Lvoncik, S., Slana, I., Kulich, P., Kralik, P., 2014. Microscopy, culture, and quantitative realtime PCR examination confirm internalization of *Mycobacteria* in plants. Appl. Environ. Microbiol. 80

797 (13), 3888-3894.

798 Karavarsamis, N., Hamilton, A.J., 2010. Estimators of annal probability of infection for quantitative microbial risk assessment. J. Water Health. 8 (2), 365 – 373.

800 Klein, W., Graesch, A.P., Izquierdo, C., 2009. Children and chores: a mixed-methods study of children's household work in Los Angeles Families. Anthropol. Work Rev. 30 (3), 98-109

802 Koplan, J.D., Dean, R.D., Swanston, W.H., Tota, B., 1978. Contaminated roof-collected rainwater as a possible cause of an outbreak of salmonellosis. J. Hyg. 81 (2), 303-309.

804 Kus, B., Kandasamy, J., Vigneswaran, S., Shon, H.K., 2010. Analysis of first flush to improve the water quality in rainwater tanks. Wat. Sci. Technol. 61 (2), 421-428.

806Lai, K.K., Stottmeier, K.D., Sherman, I.H., McCabe, W.R., 1984. Mycobacterial cervical lymphadenopathy: 807 relation of etiologic agents to age. JAMA 251 (10), 1286-1288.

808Lim, K.-Y., Hamilton, A.J., Jiang, S.C., 2015. Assessment of public health risk associated with viral

contamination in harvested urban stormwater for domestic applications. Sci. Total Environ. 523, 95-108.

811Lim, K.-Y., Jiang, S.C., 2013. Reevaluation of health risk benchmark for sustainable water practice

through risk analysis of rooftop-harvested rainwater. Water Res. 47 (20), 7273-7286.

813 Lincoln, E.M., Gilbert, L.A., 1972. Disease in children due to mycobacteria other than *Mycobacterium* 814 *tuberculosis*. Amer. Rev. Resp. Dis. 105 (5) 683-714.

815 Lopez, G.U., Gerba, C.P., Tamimi, A.H., Kitajima, M., Maxwell, S.L., Rose, J.B., 2013. Transfer efficiency

of bacteria and viruses from porous and nonporous fomites to fingers under different relative humidity

817 conditions. Appl. Environ. Microbiol. 79 (18), 5728-5734.

818Lumb, R., Stapledon, R., Scroop, A., Bond, P., Cunliffe, D., Goodwin, A., Doyle, R., Bastian, I., 2004.
Investigation of spa pools associated with lung disorders caused by *Mycobacterium avium* complex in
immunocompetent adults. Appl. Environ. Microbiol. 70 (8), 4906-4910.

821 Mayer, P.W., DeOreo, W.B., 1999. Residential end uses of water, American Water Works Association.
822 Mendez, C.B., Klenzendorf, J.B., Afshar, B.R., Simmons, M.T., Barrett, M.E., Kinney, K.A., Kirisits, M.J.,
2011. The effect of roofing material on the quality of harvested rainwater. Water Res. 45 (5), 20492059.

825 Merritt, A., Miles, R., Bates, J., 1999. An outbreak of *Campylobacter enteritis* on an island resort, north Queensland. Commun. Dis. Intell. 23 (8), 215-219.

827 Muder, R.R., Victor, L.Y., 2002. Infection due to *Legionella* species other than *L. pneumophila*. Clin. 828 Infect. Dis. 35 (8), 990-998.

829 Murrell, W., Stewart, B., 1983. Botulism in New South Wales, 1980-1981. Med. J. Aust. 1 (1), 13-17.
830 NHMRC-NRMMC., 2011. Australian drinking water guidelines 6 (Version 3.3), Natl. Health Med. Res.
831 Counc. Nat. Resour. Mag. Minist. Counc., Canberra, Australia.

832 Nolt, D. Michaels, M.G., Wald, E.R., 2003. Intrathoracic disease from nontuberculous mycobacteria in children: two cases and a review of the literature. Pediatrics 112 (5), e434-e439.

834 O'Brien, D.P., Currie, B.J., Krause, V.L., 2000. Nontuberculous mycobacterial disease in northern Australia: a case series and review of the literature. Clin. Infect. Dis. 31 (4), 958-967.

836 O'Toole, J., Keywood, M., Sinclair, M., Leder, K., 2009. Risk in the mist? Deriving data to quantify
microbial health risks associated with aerosol generation by water-efficient devices during typical
domestic water-using activities. Water Sci. Technol. 60 (11), 2913-2920.

alternative water sources: Part B-Microbial transfer efficiency during machine clothes washing and
 microbial survival turf-grass experiments, Cooperative Research Centre for Water Quality and
 Treatment.

843 O'Toole, J., Leder, K., Sinclair, M., 2008b. A series of exposure experiments–recycled water and
alternative water sources. Part A. Aerosolsizing and endotoxin experiments. , CRC for Water Quality
and Treatment, Adelaide, Australia.

846 Phin, N., Parry-Ford, F., Harrison, T., Stagg, H.R., Zhang, N., Kumar, K., Lortholary, O., Zumla, A.,

Abubakar, I., 2014. Epidemiology and clinical management of Legionnaires' disease. Lancet Infect.
Dis. 14 (10), 1011-1021.

849 Pierce, E.S., 2009. Possible transmission of *Mycobacterium avium* subspecies paratuberculosis through potable water: lessons from an urban cluster of Crohn's disease. Gut Pathog. 1 (1), 1-5.

851 Piso, R., Caruso, A., Nebiker, M., 2007. Hose as a source of *Legionella pneumonia*. A new risk factor for gardeners? J. Hosp. Infect. 67 (4), 396-397.

853 Pouillot, R., Delignette-Muller, M.-L., 2010. Evaluating variability and uncertainty in microbial quantitative risk assessment using two R packages. Int. J. Food Microbiol. 142 (3), 330-340.

855 Regli, S., Rose, J.B., Haas, C.N., Gerba, C.P., 1991. Modeling the risk from *Giardia* and viruses in drinking water. J Am. Water Works Assoc. 83 (11), 76-84.

857 Reyneke, B., Dobrowsky, P., Ndlovu, T., Khan, S., Khan, W., 2016. EMA-qPCR to monitor the efficiency
of a closed-coupled solar pasteurization system in reducing *Legionella* contamination of roofharvested rainwater. Sci. Total. Environ. 553 (X), 662-670.

860 Rice, G., Wright, J.M., Boutin, B., Swartout, J., Rodgers, P., Niemuth, N., Broder, M., 2005. Estimating the frequency of tap-water exposures to *Mycobacterium avium* complex in the US population with advanced aids. J. Toxicol. Environ. Health A . 68 (11-12), 1033-1047.

863 Rodgers, M., Blackstone, B., Reyes, A., Covert, T., 1999. Colonisation of point of use water filters by silver resistant non-tuberculous mycobacteria. J. Clinical Pathol. 52 (8), 629.

865 Sales-Ortells, H., Fernandez-Cassi, X., Timoneda, N., Dürig, W., Girones, R., Medema, G., 2014. Health

risks derived from consumption of lettuces irrigated with tertiary effluent containing norovirus. FoodRes. Int. 68, 70-77.

868 Sales-Ortells, H., Medema, G., 2012. Screening-level risk assessment of *Coxiella burnetii* (Q fever) 869 transmission via aeration of drinking water. Environ. Sci. Technol. 46 (7), 4125-4133.

870 Schets, F.M., Schijven, J.F., de Roda Husman, A.M., 2011. Exposure assessment for swimmers in 871 bathing waters and swimming pools. Water Res. 45 (7), 2392-2400.

872 Schlech III, W.F., Gorman, G.W., Payne, M.C., Broome, C.V., 1985. Legionnaires' disease in the

873 Caribbean: an outbreak associated with a resort hotel. Arch. Intern. Med. 145 (11), 2076.

874 Schoen, M.E., Ashbolt, N.J., 2011. An in-premise model for *Legionella* exposure during showering events. 875 Water Res. 45 (18), 5826-5836.

876 Schoen, M.E., Garland, J., 2015. Review of pathogen treatment reductions for onsite non-potable reuse

877 of alternative source waters. Microbial Risk Anal. (online early).

- 878 Schoen, M.E., Xue, X., Hawkins, T.R., Ashbolt, N.J., 2014. Comparative human health risk analysis of
 coastal community water and waste service options. Environ. Sci. Technol. 48 (16), 9728-9736.
 880 Shuval, H., Lampert, Y., Fattal, B., 1997. Development of a risk assessment approach for evaluating
 wastewater reuse standards for agriculture. Wat. Sci. Technol. 35 (11), 15-20.
- wastewater reuse standards for agriculture. Wat. Oct. Feelinol. 35 (Fr), 13-20.
 882 Simmons, G., Jury, S., Thornley, C., Harte, D., Mohiuddin, J., Taylor, M., 2008. A Legionnaires' disease
 outbreak: a water blaster and roof-collected rainwater systems. Water Res. 42 (6-7), 1449-1458.
 884 Simmons, G., Smith, J., 1997. Roof water probable source of *Salmonella* infections. New Zealand Public
 Health Report. 4 (1), 5.
- 886 Sinclair, M., Roddick, F., Nguyen, T., O'Toole, J., Leder, K., 2016. Measuring water ingestion from spray 887 exposures. Water Res. 99, 1-6.
- 888 Soller, J.A., Eftim, S.E., Warren, I., Nappier, S.P., 2016. Evaluation of microbiological risks associated with direct potable reuse. Microb. Risk Anal. In press. <u>http://dx.doi.org/10.1016/j.mran.2016.08.003</u>
- 890 Solomon, E.B., Yaron, S., Matthews, K.R., 2002. Transmission of *Escherichia coli* O157: H7 from
- contaminated manure and irrigation water to lettuce plant tissue and its subsequent internalization.
 Appl. Environ. Microbiol. 68 (1), 397-400.
- 893 Steed, K.A., Falkinham 3rd, J.O., 2006. Effect of growth in biofilms on chlorine susceptibility of 894 *Mycobacterium avium* and *Mycobacterium intracellulare*. Appl. Environ. Microbiol. 72 (6), 4007-4011.
- 895 Taylor, R.H., Falkinham 3rd, J.O., Norton, C.D., LeChevallier, M.W., 2000. Chlorine, chloramine, chlorine
 dioxide, and ozone susceptibility of *Mycobacterium avium*. Appl. Environ. Microbiol. 66 (4), 1702 1705.
- 898 Tebruegge, M., Pantazidou, A., MacGregor, D., Gonis, G., Leslie, D., Sedda, L., Ritz, N., Connell, T.,
 Curtis, N., 2016. Nontuberculous mycobacterial disease in children—epidemiology, diagnosis &
 management at a tertiary center. PloS One 11 (1), e0147513.
- 901 Thomson, R.M., Carter, R., Tolson, C., Coulter, C., Huygens, F., Hargreaves, M., 2013. Factors
 associated with the isolation of Nontuberculous mycobacteria (NTM) from a large municipal water
 system in Brisbane, Australia. BMC Microbiol. 13, 89.
- 904 USEPA (2011a) Exposure Factors Handbook, EPA/600/R-090/052F. National Center for Environmental 905 Assessment, Office of Research and Development, Washington, DC.
- 906 USEPA (2011b) Recreational Water Criteria.
- 907 Villarreal, E.L., Dixon, A., 2005. Analysis of a rainwater collection system for domestic water supply in 908 Ringdansen, Norrkoping, Sweden. Build. Environ. 40 (9), 1174-1184.
- 909Waddell, L., Rajić, A., Stark, K. and McEWEN, S., 2015. The zoonotic potential of *Mycobacterium avium*910 ssp. *paratuberculosis*: a systematic review and meta-analyses of the evidence. Epidemil. Infection.
 911 143 (15), 3135-3157.
- 912 Wallace, R.J., Iakhiaeva, E., Williams, M.D., Brown-Elliott, B.A., Vasireddy, S., Vasireddy, R., Lande, L.,
- 913 Peterson, D.D., Sawicki, J., Kwait, R., 2013. Absence of *Mycobacterium intracellulare* and presence 914 of *Mycobacterium chimaera* in household water and biofilm samples of patients in the United States
- 915 with Mycobacterium avium complex respiratory disease. J. Clin. Microbiol. 51 (6), 1747-1752.
- 916Wei, J., Jin, Y., Sims, T., Kniel, K.E., 2011. Internalization of murine norovirus 1 by Lactuca sativa during 917 irrigation. Appl. Environ. Microbiol. 77 (7), 2508-2512.
- 918 Whiley, H., Keegan, A., Fallowfield, H., Bentham, R., 2014. Detection of *Legionella*, *L. pneumophila* and
 Mycobacterium avium complex (MAC) along potable water distribution pipelines. Int. J. Environ. Res.
 Public Health. 11 (7), 7393-7405.
- 921WHO., 2004. Guidelines for drinking water quality, World Health Organization, Geneva, Switzerland. 922Wolinsky, E., 1995. Mycobacterial lymphadenitis in children: a prospective study of 105 nontuberculous 923 cases with long-term follow-up. Clin. Infect. Dis. 20(4): 954-963.
- 924 Yajko, D., Nassos, P., Sanders, C., Gonzalez, P., Reingold, A., Horsburgh, C., Hopewell, P., Chin, D.,
- Hadley, W., 1993. Comparison of four decontamination methods for recovery of *Mycobacterium*
- 926 *avium* complex from stools. J. Clin. Microbiol. 31 (2), 302-306.

Table 1 927

928 Monte Carlo simulation input parameters for ingestion scenarios (for MAC only)

Parameter	Symbol	Unit	Value	Distribution	Source
Child population parameters					
Portion of total age 1-12 MAC-	P _{attr,i}	Proportion	0.60	Point	(Lai et al. 1984)
attributable lymphadenitis cases ages 1-	- au,i				(,
2					
Portion of total age 1-12 MAC-	P _{attr,i}	Proportion	0.27	Point	(Lai et al. 1984)
attributable lymphadenitis cases ages 3-	utti,/				(
5					
Portion of total age 1-12 MAC-	P _{attr,i}	Proportion	0.13	Point	(Lai et al. 1984)
attributable lymphadenitis cases ages 6-	any	·			, , , , , , , , , , , , , , , , , , ,
12					
Drinking water					
Intake rate (general population)	V _{ing,dw}	L per day	$\mu = -0.529, \sigma = 0.882$	Lognormal ^a	(USEPA 2011a)
Intake rate (children 1-2 years)	V _{ing,dw}	L per day	$\mu = -1.578, \sigma = 0.824$	Lognormal ^a	(USEPA 2011a)
Intake rate (children 3-5 years)	V _{ing,dw}	L per day	$\mu = -1.457$, $\sigma = 0.823$	Lognormal ^a	(USEPA 2011a)
Intake rate (children 6-12 years)	V _{ing,dw}	L per day	$\mu = -1.284, \sigma = 0.897$	Lognormal ^a	(USEPA 2011a)
Exposure frequency	n _{dw}	-	365	Point	Assumption
Log removals due to filtration	L	logs	Min=0.4, Max=2	Uniform	(Jordan 2008)
Toilet flushing					х <i>у</i>
Volume ingested	$V_{ing,t}$	mL	Min = 0.01, Max = 0.3	Uniform	(NRMMC-EPHC-AHMC 2006,
-	.				Schoen et al. 2014)
Toilet flushes per day	n _t	Flushes per day	5	Point	(Mayer and DeOreo 1999)
Showering					
Volume ingested	V _{ing,sh}	mL	Min = 0.058, Max = 1.9	Uniform	(Ahmed et al. 2010)
Showers per year	n _{sh}	Number per year	365	Point	Assumption
Garden hosing					
Volume ingested	V _{ing,gh}	mL	$Min = 2 \times 10^{-9}$, $Max = 1.1 \times 10^{-3}$	Uniform	(Ahmed et al. 2010, NRMMC-
					EPHC-AHMC 2006, Schoen e
					al. 2014)
Hosing events per year	n _{gh}	Number per year	90	Point	(NRMMC-EPHC-AHMC 2006)
Car washing					
Volume ingested	V _{ing,cw}	mL	Min = 0.06, Max = 3.79	Uniform	(Sinclair et al. 2016)
Car washing events per year	n _{cw}	Number per year	12	Point	(Villarreal and Dixon 2005)
Pool top-up and bathing					
Dilution Factor (RHRW: sterile water)	D	%	10	Point	Assumption
			50		
			90		
			100 (bathing only)		

Volume ingested per swim (general population)	V _{ing,sw}	mL	μ = 2.92 , σ = 1.43	Lognormal	(Dufour et al. 2006, Schoen et a 2014)		
Volume ingested per swim (children age 1-12) or bath (children age <6)	V _{ing,sw}	mL	r = 0.81, λ = 63	Gamma	(Schets et al. 2011)		
Swims per year (general population)	n _{sw}	Number per year	Average 21 Upper bound 72	Point	(Schets et al. 2011) ^b		
Swims per year (children age 1-12)	n _{sw}	Number per year	Average 32 Upper bound 122	Point	(Schets et al. 2011) ^b		
Baths per year	n _{bath}	Number per year	365	Point	Assumption		
Clothes washing	· ·bau						
Volume ingested	V _{ina.cw}	mL	0.01	Point	(NRMMC-EPHC-AHMC 2006)		
Number of times clothes are washed with rainwater per year	n _{cw}	Number per year	100	Point	(NRMMC-EPHC-AHMC 2006)		
Produce consumption							
Volume irrigated water retained on lettuce	V _R	mL per g	$\mu = 0.108$, $\sigma = 0.019$	Normal, truncated at 0	(Shuval et al. 1997)		
Internalized fraction of MAC in irrigation water	F _{int}	Proportion	Min=1.13 × 10 ⁻⁵ , Max=9.49 × 10 ⁻⁴	Uniform	(Kaevska et al. 2014)		
In-field decay on surface of plant	K _{f,s}	d-1	-0.0484	Point	(Cook et al. 2013)		
Log reductions due to lettuce washing	W	Logs	$\mu = 0.3, \sigma = 0.1$	Normal, truncated at 0	(Holvoet et al. 2014)		
Time in field between irrigation and harvest	t _f	day	2	Point	(Barker et al. 2013)		
Intake rate (general population)	1	g/kg-day	$\mu = -2.30, \sigma = 1.64$	Lognormal	(Barker et al. 2013)		
Intake rate (1-2 years)	1	g/kg-day	$\mu = -3.388, \sigma = 1.727$	Lognormal	(USEPA 2011a)		
Intake rate (3-5 years)	1	g/kg-day	$\mu = -2.905, \sigma = 1.694$	Lognormal	(USEPA 2011a)		
Intake rate (6-12 years)		g/kg-day	$\mu = -2.860, \sigma = 1.475$	Lognormal	(USEPA 2011a)		
Body weight (general)	B _{gen}	kg	$\mu = 4.136$, $\sigma = 0.434$	Lognormal	Modelled parameters based on (ABS 1998, ABS 2016, CSIRO 2008)		
Body weight (1-2 years)	B ₁₋₂	kg	$\mu = 2.522$, $\sigma = 0.152$	Lognormal	(USEPA 2011a)		
Body weight (3-5 years)	B ₃₋₅	kg	$\mu = 2.902, \sigma = 0.206$	Lognormal	(USEPA 2011a)		
Body weight (6-12 years)	B ₆₋₁₂	kg	$\mu = 3.407$, $\sigma = 0.323$	Lognormal	(USEPA 2011a)		
Proportion who consume lettuce (general population)	F _{cl}	Proportion	0.53	Point	(USEPA 2011a)		
Proportion who consume lettuce (1-2 years)	F _{cl}	Proportion	0.21	Point	(USEPA 2011a)		
Proportion who consume lettuce (3-5 years)	F _{cl}	Proportion	0.29	Point	(USEPA 2011a)		
Proportion who consume lettuce (6-12 vears)	F _{cl}	Proportion	0.37	Point	(USEPA 2011a)		
Lettuce consumption events per year	n	Days per year	365	Point	USEPA intake rates reported as normalized per day		

^aLognormal parameters mean, standard deviation (μ , δ) calculated from population (normal) parameters (\bar{x} , s) using standard formulae as follows: $\mu = \ln(\bar{x}^2/(s^2 + \bar{x}^2)^{1/2}), \delta = [\ln(1 + (s^2/\bar{x}^2))]^{1/2}, \text{ where } \bar{x} \text{ is the sample mean and } s^2 \text{ is the sample standard deviation.}$ ^b Estimates from Schets et al. modified based on a 9-month Queensland swim season.

Table 2. 949

Monte Carlo simulation input parameters for inhalation scenarios. 950

Parameter	Symbol	Unit	Value	Distribution	Source		
Breathing rate, light activity, breathing	В	m ³ per min	0.013-0.017	Uniform	(USEPA 2011a)		
cycle period 8 s and 1 L tidal volume							
Deposition efficiency (diameter)	DE_i	Proportion		Uniform	(Heyder et al. 1986)		
1			Min = 0.23, Max =0.25				
2			Min = 0.4, Max =0.53				
3			Min = 0.36, Max =0.62				
4			Min = 0.29, Max =0.61				
5			Min = 0.19, Max =0.52				
6			Min = 0.1, Max =0.4				
7			Min = 0.06, Max =0.29				
8			Min = 0.03, Max =0.19				
9			Min = 0.01, Max =0.12				
10			Min = 0.01, Max =0.06				
Toilet flushing							
Toilet flushes per day	f _t	Flushes per day	5	Point	(Mayer and DeOreo 1999)		
Time in bathroom after flush	t_t	Min per flush	Min = 1, Max = 5	Uniform	Lim et al. (2015)		
Concentration of aerosols of diameter i:	C _{aer,i}	# aerosols per cm ³ of	μ = -1.246, σ = 1.885	Lognormal ^a	(O'Toole et al. 2009)		
2.5		air					
Showering							
Shower duration	t _{sh}	min per day	15	Point	(Schoen and Ashbolt 2011)		
Showers per year	n _{sh}	Showers per year	365	Point	Assumption		
Concentration of aerosols of diameter i:	C _{aer,i}	# aerosols per cm ³ of		Lognormal	(O'Toole et al. 2009)		
1.5		air					
2.5			μ = 3.718, σ = 0.296				
reathing rate, light activity, breathing vcle period 8 s and 1 L tidal volume eposition efficiency (diameter) oilet flushing bilet flushes per day ime in bathroom after flush oncentration of aerosols of diameter <i>i</i> . 2. howering hower duration howers per year oncentration of aerosols of diameter <i>i</i> . 1. 2. 4. tarden hosing osing duration osing events per year oncentration of aerosols of diameter <i>i</i> . 1. 2. 4. tarden hosing oncentration of aerosols of diameter <i>i</i> . 1. 2. 4. 4. 4.			μ = 3.699, σ = 0.170				
8			μ = 5.549, σ = 0.348				
reathing rate, light activity, breathing role period 8 s and 1 L tidal volume eposition efficiency (diameter) bilet flushing bilet flushes per day me in bathroom after flush oncentration of aerosols of diameter <i>i</i> . 2. howering hower duration howers per year oncentration of aerosols of diameter <i>i</i> . 1. 2. arden hosing bilet flushing bilet flushes per year oncentration of aerosols of diameter <i>i</i> . 1. 2. 4. 2. 5. 5. 5. 5. 5. 5. 5. 5. 5. 5. 5. 5. 5.			μ = 6.185, σ = 0.309				
-							
Hosing duration	t _{gh}	min	7	Point	(Ahmed et al. 2010)		
Hosing events per year	n _{gh}	Number per year	90	Point	(NRMMC-EPHC-AHMC 2006		
Concentration of aerosols of diameter <i>i</i> :	C _{aer,i}	# aerosols per cm ³ of		Lognormal	(O'Toole et al. 2009)		
		air					
1.5			μ = 5.728, σ = 0.274				
2.5			μ = 4.949, σ = 0.333				
4.5			$\mu = 3.047, \sigma = 0.586$				
8			μ = -2.451=, σ=1.579				
Car washing							

Car washing duration	t _{cw}	min	10	Point	(O'Toole et al. 2008b)
Car washing events per year	n _{cw}	Number per year	12	Point	(Villarreal and Dixon 2005)
Concentration of aerosols of diameter i:	C _{aer,i}	# aerosols per cm ³ of		Lognormal	(O'Toole et al. 2009)
1.5		air			
2.5			μ = 6.187, σ = 0.476		
4.5			μ = 4.665, σ = 0.420		
8			μ = 1.742, σ = 0.591		
			$\mu = -1.551, \sigma = 0.833$		
Pool top-up					
Partitioning coefficient- L. pneumophila	P_{LP}	L per m ³	Min = 2.2×10 ⁻⁸ , Max = 1.1 × 10 ⁻⁵	Uniform	(Hines et al. 2014)
Partitioning coefficient- MAC	P _{MAC}	L per m ³	Min = 1.0×10^{-4} , Max = 5.3 $\times 10^{-3}$	Uniform	(Glazer et al. 2007)
Dilution Factor (RHRW:sterile water)	D	%	10	Point	Assumption
· · · · · · · · · · · · · · · · · · ·			50		•
			90		
Pool exposure time per swim (general)	tpool	min	$\mu = 4.082, \sigma = 0.564$	Lognormal	(Schets et al. 2011)
Swims per year (general population)	n _{sw}	Number per year	Average 21 Upper bound 72	Point	(Schets et al. 2011) ^b

^a Lognormal parameters mean, standard deviation (μ , δ) calculated from population (normal) parameters (\bar{x} , s) using standard formulae as follows: $\mu = \ln(\bar{x}^2/(s^2 + \bar{x}^2)^{1/2})$, $\delta = [\ln(1 + (s^2/\bar{x}^2))]^{1/2}$, where \bar{x} is the sample mean and s^2 is the sample standard deviation. ^b Estimates from Schets et

al. modified based on a 9-month Queensland swim season.

Table 3.

~	~	~
9	6	9

Parameter	Symbol	Unit	Value	Distribution	Source	
L. pneumophila						
Dose response parameter for L.	r	Unitless	$\mu = -2.934, \sigma = 0.488$	Lognormal ^a	(Armstrong and Haas 2007	
pneumophila, infection endpoint						
MAC						
Dose response parameter for M.	r	Unitless	μ = -13.742, σ = 0.208	Lognormal	(Hamilton et al. 2017)	
avium- pulmonary infection						
(subclinical infection endpoint)						
Conversion factor from intravenous	С	Unitless	500	Point	(Hamilton et al. 2017)	
to inhalation route for pulmonary						
infection model	~	Unitless	0.201	Point	(Hamilton at al. 2017)	
Dose response parameters for	α	Unitiess	0.201 1.15 × 10 ⁻⁶	Point	(Hamilton et al. 2017)	
discominated infaction						
disseminated infection	β	Linitiess		Lognormal	(Hamilton et al. 2017)	
Dose response models for cervical	β r	Unitless	$\mu = -19.006, \sigma = 1.008$	Lognormal	(Hamilton et al. 2017)	
Dose response models for cervical lymphadenitis in children	r		μ = -19.006, σ = 1.008	-	, , , , , , , , , , , , , , , , , , ,	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	
Dose response models for cervical lymphadenitis in children ^a Lognormal parameters mean, sta	r ndard deviation	(μ, δ) calculated	μ = -19.006, σ = 1.008 from population (normal) para	meters (\bar{x}, s) using	standard formulae as	

Table 4.

987 Monte carlo simulation input parameters for pathogen concentrations.

CONCENTRATIONS IN RHRW					
M. avium	$C_{RHRW,MA}$	# per L	μ = 0.723, σ = 4.349	Lognormal ^a	(Hamilton et al. 2016)
M. intracellulare	$C_{RHRW,MI}$	# per L	μ = 6.720, σ = 2.410	Lognormal	(Hamilton et al. 2016)
L. pneumophila, positive samples only	C _{RHRW,LP}	# per L	μ = 8.080, μ = 0.745	Lognormal	(Hamilton et al. 2016)
Probability of L. pneumophila	Pcontam	Fraction	n = 134, p = 0.03	Binomial	(Hamilton et al. 2016)
occurrence					
Recovery efficiency	R	Fraction	0.84	Point	(Hamilton et al. 2016)
^a Lognormal parameters mean, sta					
follows: $\mu = \ln(\bar{x}^2/(s^2 + \bar{x}^2)^{1/2}), \delta = [\ln(s^2/(s^2 + \bar{x}^2)^{1/2})]$	$1+(s^2/\bar{x}^2))]^{1/2}$	2 , where \bar{x} is the	e sample mean and s ² is th	e sample standard de	eviation.

Table 5

Sensitivity analysis with Spearman rank correlation coefficients for ingestion risks. Risks for endpoints of children with cervical lymphadenitis and severe immune deficiency with disseminated infection are shown.

Scenario	Population			Parame	ter (Spe	arman rank correl	ation co	efficien	t)	
		В	C_{MA}	C _{MI}	F _{int}	Ι	L	r	Ving	W
Drinking water- filtration	Children		0.16	0.73				0.34	0.15 (1 to 2) 0.08 (3 to 5) 0.06 (6 to 12)	
	Immune-comp.		0.17	0.76			-0.36		0.30	
Drinking water- no filtration	Children		0.18	0.80				0.37	0.16 (1 to 2) 0.08 (3 to 5) 0.05 (6 to 12)	
•	Immune-comp.		0.18	0.83					0.33	
Lettuce	Children	0.02 (1 to 2) 0.03 (3 to 5) 0.03 (6 to 12)	0.16	0.74	-0.01	0.20 (1 to 2) 0.20 (3 to 5) 0.15 (6 to 12)		0.34	0.05*	-0.08
	Immune-comp.	0.14	0.16	0.71	0.002	0.54			0.05*	-0.08
Shower	Children (6-12 y)		0.17	0.78				0.36	0.26	
	Immune-comp.		0.18	0.85					0.29	
Bathing	Children (1-5 y)		0.16	0.71				0.33	0.46	
Hose	Immune-comp.		0.18	0.82					0.33	
Car	Immune-comp.		0.19	0.84					0.31	
Toilet	Children (>3 y)		0.17	0.78				0.37	0.27	
	Immune-comp.		0.19	0.85					0.29	
Clothes washing	Immune-comp.		0.19	0.90						
Pool, D = 10%, n = Avg.	Children		0.16	0.71				0.33	0.46	
	Immune-comp.		0.17	0.76					0.51	
Pool, D = 50%, n = Avg.	Children		0.16	0.71				0.33	0.46	
	Immune-comp.		0.17	0.75					0.50	
Pool, D = 90%, n = Avg.	Children		0.16	0.71				0.33	0.46	
	Immune-comp.		0.17	0.75					0.50	
Pool, D = 10%, n = High	Children		0.15	0.70				0.33	0.46	

	Immune-comp.	0.16	0.75		0.50
Pool, $n = 50\%$, $n = High$	Children	0.16	0.71	0.33	0.45
	Immune-comp.	0.16	0.76		0.50
Pool, $D = 90\%$, $n = High$	Children	0.16	0.71	0.33	0.46
	Immune-comp.	0.17	0.75		0.50

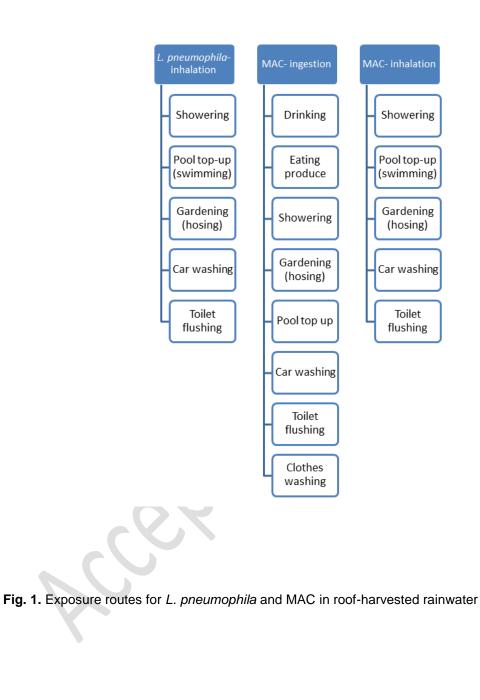
1015 *For the lettuce scenario, the coefficient for volume of water retained by lettuce (V_R) is shown

1016 **Table 6**

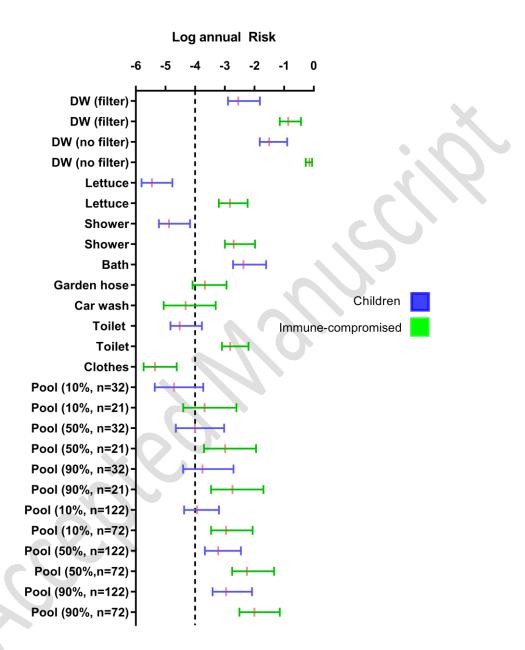
1017 Sensitivity analysis with Spearman rank correlation coefficients for inhalation risk scenarios. Risks for endpoints of pulmonary infection in healthy 1018 populations with *L. pneumophila* or MAC are shown.

Scenario	Population					P	arameter (Spearn	nan ran	k correla	tion coeff	icient)					
		В	Caer1.5	Caer2.5	Caer4.5	C _{aer8}	C _{RHRW}	Сма	Смі	DE1.5	DE _{2.5}	DE4.5	DE ₈	Р	Pcontam	r	t
Toilet flushing	LP	0.04		0.80			0.31				0.05				0.26	0.20	0.18
	MAC	0.03		0.58				0.14	0.65		0.04					0.06	0.13
Shower	LP	0.06	0.002	0.003	0.10	0.16	0.59			0.002	-0.002	0.05	0.19		0.46	0.39	
	MAC	0.03	-0.002	-0.005	0.05	0.07		0.19	0.88	0.005	-0.005	0.03	0.09			0.09	
Garden hose	LP	0.06	0.03	0.13	0.16	0.003	0.60			0.03	0.05	0.05	-0.002		0.47	0.39	
	MAC	0.03	0.02	0.07	0.08	-0.001		0.19	0.88	0.01	0.02	0.03	-0.005			0.08	
Car wash	LP	0.06	0.12	0.15	0.06	0.07	0.61			0.07	0.04	0.01	0.03		0.46	0.39	
	MAC	0.04	0.06	0.08	0.03	0.04		0.19	0.88	0.03	0.02	0.02	0.02			0.08	
Pool, D=10%, n=Avg.	LP	0.05					0.45							0.50	0.36	0.30	0.34
	MAC	0.04						0.19	0.82					0.29		0.08	0.20
Pool, D=50%, n=Avg.	LP	0.05					0.46							0.50	0.36	0.29	0.34
	MAC	0.03						0.18	0.81					0.29		0.08	0.21
Pool, D=90%, n=Avg.	LP	0.04					0.45							0.50	0.37	0.30	0.34
	MAC	0.03						0.18	0.81					0.29		0.08	0.21
Pool, D=10%, n=High	LP	0.05					0.45							0.50	0.36	0.29	0.34
	MAC	0.03						0.18	0.82					0.30		0.08	0.21
Pool, D=50%, n=High	LP	0.05					0.45							0.50	0.36	0.30	0.34
	MAC	0.02						0.18	0.82					0.29		0.08	0.21
Pool, D=90%, n=High	LP	0.05					0.45							0.50	0.36	0.30	0.34
	MAC	0.03						0.17	0.81					0.30		0.07	0.21

- 1019 Fig1



1030



1031

1032

Fig. 2. Annual cervical lymphadenitis (children) or disseminated infection (immune-compromised populations) risks for ingestion of *Mycobacterium avium* complex through food or water exposure scenarios. Median and 95% confidence intervals and scenario analysis for various pool dilution levels (D = 10%, 50%, or 90% RHRW: sterile water), with or without drinking water under-sink point of use filtration (filtration/no filtration), and swimming frequency assumptions (32 or 122 swims per year for children and

1038 21 or 72 swims per year for a general population) are shown.

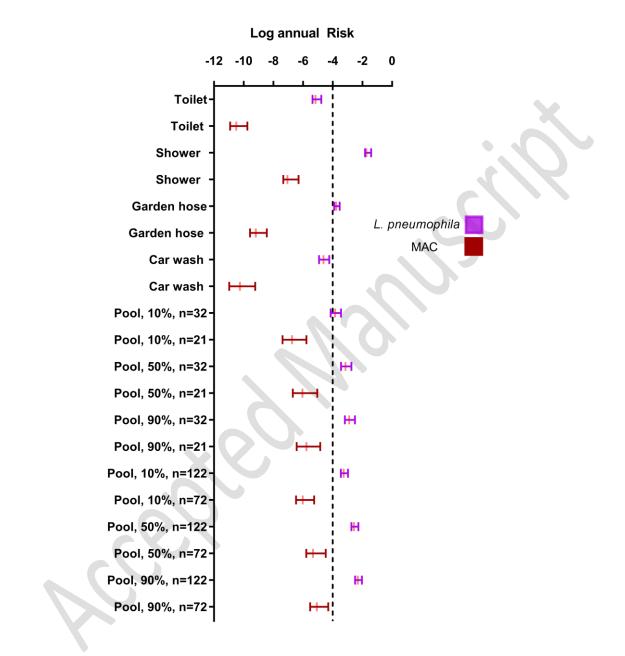
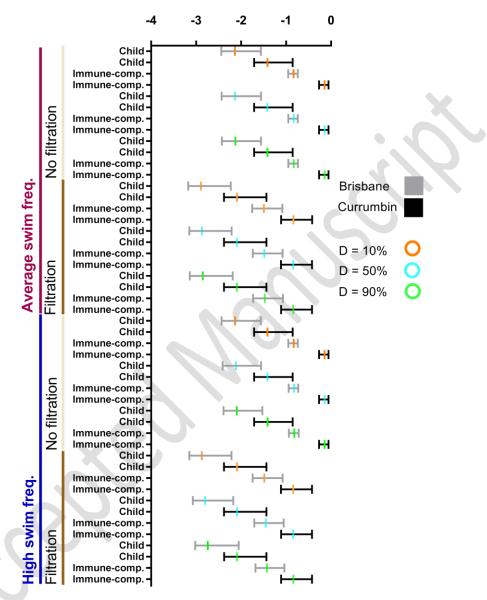


Fig. 3. Annual infection risks for inhalation of *Mycobacterium avium* complex or L. *pneumophila* for
 various exposure scenarios. Median and 95% confidence intervals and scenario analysis for various pool
 dilution levels (D = 10%, 50%, or 90% RHRW: sterile water) and swimming assumptions (21 or 72 swims
 per for a general population) are shown.

Log annual Risk



1049

1050

Fig. 4. Total annual risks from all activities for cervical lymphadenitis in children or disseminated infection
 in immune-compromised populations via ingestion. Median and 95% confidence intervals shown.
 Scenario analysis for various pool dilution levels (D = 10%, 50%, or 90% RHRW: sterile water), with or
 without drinking water under-sink point of use filtration (filtration/no filtration), and swimming frequency
 assumptions (32 or 122 swims per year) are shown.

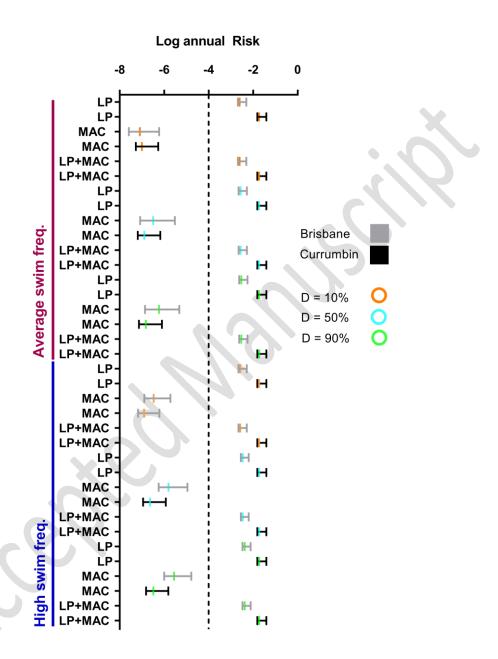


Fig. 5. Total annual pulmonary infection risks from all activities via inhalation of *Mycobacterium avium* complex (MAC), *L. pneumophila* (LP), or both organisms (LP+MAC). Median and 95% confidence
 intervals are shown. Scenario analysis for various pool dilution levels (D = 10%, 50%, or 90% RHRW:
 sterile water) and swimming assumptions (32 or 122 swims per year) are shown.